



PCT

WORLD INTELLECTUAL PROPERTY ORGANIZATION  
International Bureau

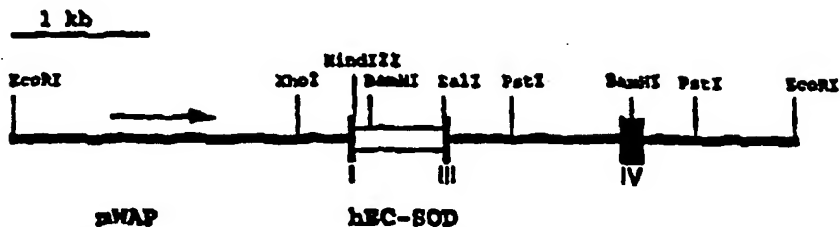
## INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification <sup>5</sup> : C12N 15/00, A01K 67/027, C12N 9/02, 15/53, 5/10, A23C 9/20		A1	(11) International Publication Number: <b>WO 95/00637</b>
			(43) International Publication Date: 5 January 1995 (05.01.95)
(21) International Application Number: PCT/IB94/00181		(81) Designated States: AM, AU, BB, BG, BR, BY, CA, CN, CZ, CZ (Utility model), DE, DE (Utility model), DK, DK (Utility model), FI, FI (Utility model), GE, HU, JP, KE, KG, KP, KR, KZ, LK, LV, MD, MG, MN, MW, NO, NZ, PL, RO, RU, SD, SI, SK, SK (Utility model), TJ, TT, UA, US, UZ, VN, European patent (AT, BE, CH, DE, DK, ES, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, ML, MR, NE, SN, TD, TG).	
(22) International Filing Date: 24 June 1994 (24.06.94)		Published With international search report.	
(30) Priority Data: 0753/93 24 June 1993 (24.06.93) DK			
(71) Applicant (for all designated States except US): SYMBICOM AKTIEBOLAG [SE/SE]; Tvistevägen 48, P.O. Box 1451, S-907 36 Umeå (SE).			
(72) Inventor; and (75) Inventor/Applicant (for US only): HANSSON, Lennart [SE/SE]; Björkvägen 50, S-902 40 Umeå (SE).			
(74) Agent: PLOUGMANN & VINGTOFT A/S; Sankt Annæ Plads 11, P.O. Box 3007, DK-1021 Copenhagen K (DK).			

(54) Title: TRANSGENIC PRODUCTION OF EC-SOD

## (57) Abstract

The present invention relates to a mammalian expression system comprising a DNA sequence encoding a polypeptide having the amino acid sequence shown in SEQ ID NO:2 or a variant thereof which has a biological activity of human EC-SOD. The mammalian expression system is preferably expressed in a non-human mammal selected from the group consisting of rabbits, mice, rats, goats, sheep, pigs, lama, camels and bovine species. The invention includes expression systems, wherein the variant has a reduced heparin affinity as compared to human EC-SOD type C such as the variants T216, T215, T213, T209, SAT216, SRT216, SA219, SA220 and SA211-213, and expression systems wherein the variant has an increased heparin affinity as compared to human EC-SOD type C such as the variants SA216, SA216/218 and SA216/220. Within the scope of the invention is also DNA fragments, hybrid genes, expression vectors, cells, methods for producing a transgenic non-human mammal capable of expressing EC-SOD as defined above, and non-human mammals expressing EC-SOD.



# Best Available Copy

## FOR THE PURPOSES OF INFORMATION ONLY

Codes used to identify States party to the PCT on the front pages of pamphlets publishing international applications under the PCT.

AT	Austria	GB	United Kingdom	MR	Mauritania
AU	Australia	GE	Georgia	MW	Malawi
BB	Barbados	GN	Guinea	NE	Niger
BE	Belgium	GR	Greece	NL	Netherlands
BF	Burkina Faso	HU	Hungary	NO	Norway
BG	Bulgaria	IE	Ireland	NZ	New Zealand
BJ	Benin	IT	Italy	PL	Poland
BR	Brazil	JP	Japan	PT	Portugal
BY	Belarus	KE	Kenya	RO	Romania
CA	Canada	KG	Kyrgyzstan	RU	Russian Federation
CF	Central African Republic	KP	Democratic People's Republic of Korea	SD	Sudan
CG	Congo	KR	Republic of Korea	SE	Sweden
CH	Switzerland	KZ	Kazakhstan	SI	Slovenia
CI	Côte d'Ivoire	LI	Liechtenstein	SK	Slovakia
CM	Cameroon	LK	Sri Lanka	SN	Senegal
CN	China	LU	Luxembourg	TD	Chad
CS	Czechoslovakia	LV	Latvia	TG	Togo
CZ	Czech Republic	MC	Monaco	TJ	Tajikistan
DE	Germany	MD	Republic of Moldova	TT	Trinidad and Tobago
DK	Denmark	MG	Madagascar	UA	Ukraine
ES	Spain	ML	Mali	US	United States of America
FI	Finland	MN	Mongolia	UZ	Uzbekistan
FR	France			VN	Viet Nam
GA	Gabon				

# Best Available Copy

1

## TRANSGENIC PRODUCTION OF EC-SOD

The present invention relates to a mammalian expression system comprising a DNA sequence encoding human extra cellular superoxide dismutase (EC-SOD) or encoding a variant of said protein having the superoxide dismutating property of native extracellular superoxide dismutase by means of production in transgenic non-human mammals, and to a method of producing a transgenic mammal capable of expressing EC-SOD or a variant thereof. The present invention furthermore relates to a genomic DNA sequence advantageously used in the production of recombinant human EC-SOD or a variant thereof.

Organisms living in the presence of oxygen have been forced to develop a number of protective mechanisms against toxic oxygen reduction metabolites, such as superoxide radicals, which are formed in connection with a variety of biological oxidations. The protective factors include superoxide dismutases (SOD) which dismutate the superoxide radical and are found in relatively constant amounts in mammalian cells and tissue. The secretory extracellular superoxide dismutase (EC-SOD) is one of three different SOD isoenzymes present in mammals [1]. It is the major SOD isoenzyme in plasma, lymph and synovial fluid [2-5], but exists primarily in the interstitial space of tissues [6-8]. The other two isoenzymes are the dimeric CuZn-SOD, which is an intracellular cytosolic enzyme and the tetrameric Mn-SOD, which is found in the mitochondrial matrix [9-11]. The human EC-SOD cDNA has been isolated and sequenced [12] and the recombinant protein has been produced in mammalian cells (WO 87/01387)

The DNA sequence shown in SEQ ID NO:1 is the part of the cDNA sequence corresponding to amino acid residues 1-222. EC-SOD is a secretory protein and the complete cDNA also encodes an 18 amino acids long signal sequence which is absent in mature and recombinant EC-SOD. [12] As deduced from the cDNA sequence, the subunit molecular weight of the mature enzyme is 24,200. The exact size of the carbohydrate substituent is not





## Best Available Copy

2

known, but the apparent molecular weight on gel chromatography is 140-150 kDa. EC-SOD is a tetrameric Cu and Zn containing glycoprotein. On SDS-PAGE electrophoresis the subunits display a molecular weight of 30-32 kDa. The sequence  
5 contains one glycosylation site Asn-89. The tetramers contain 4 Cu and 4 Zn atoms, in that each subunit binds one copper and one zinc atom. The active site, which contains the metal atoms, is homologous to the active site of the intracellular CuZn SODs.

- 10 The part of the EC-SOD constituting amino acid sequence 1-96 is contemplated to be involved in the formation of oligomers of the polypeptide whereas the part of EC-SOD constituting amino acid sequence 97-193 is contemplated to comprise the active site of the enzyme.
- 15 The glycan of the native protein has not been subject for any thorough investigation but the mature enzyme binds to the lectins concanavalin A, wheat germ lectin and lentil lectin. A fundamental and distinguishing property of EC-SOD is its affinity for some glycosaminoglycans such as heparin and  
20 heparan sulfate [1, 3, 4, 16, 17]. The latter, which exists in the glycocalyx of cell surfaces and in the connective tissue matrix, is the important physiological ligand of EC-SOD. Because of this affinity, EC-SOD forms in the vasculature an equilibrium between the plasma phase and the glycocalyx of the endothelium [3, 4, 15, 18]. In tissues, virtually  
25 all EC-SOD exists anchored to heparan sulfate in the interstitial space and in the cell surfaces [8].

Upon intravenous injection, EC-SOD rapidly binds to the surface of the endothelium and it displays a long half-life  
30 (15-20 hours) in the vasculature [8, 15]. When administered parenterally, CuZn-SOD has been shown to exert a host of therapeutic actions [21-25]. The experience with EC-SOD is in comparison small [26-33] but this isoenzyme seems to be even more potent. The high efficacy is apparently related to the  
35 special pharmacokinetic properties of the enzyme.



The heparin-binding domain of the enzyme is located to the positively charged carboxyterminal end [19]. This domain is easily proteolysed [20] resulting *in vivo* in EC-SOD forms with reduced and absent heparin affinities [8].

- 5 EC-SOD isolated from tissues and plasma is heterologous with regard to heparin affinity, and can upon chromatography on Heparin-Sepharose be divided into three subclasses;

- A which does not bind,  
B with intermediate affinity and  
10 C with relatively strong heparin affinity.

*In vivo* the correlate of the heparin affinity is binding to heparan sulfate proteoglycan, which occurs on cell surfaces and in the interstitial connective tissue.

- Upon injection of heparin, bound enzyme is released to plasma  
15 because it binds to the heparin instead of the heparan sulfate.

- EC-SOD Cs appear to bind to the surface of most cell-types in the body, notable exceptions are erythrocytes and neutrophilic leukocytes. Binding to various *E. coli* strains could  
20 not be demonstrated. This binding pattern suggests that EC-SOD C has the potential to protect most normal cells in the body, without protecting microorganisms lacking affinity and without much interfering with superoxide radicals produced at the surfaces of activated neutrophil leukocytes.

- 25 In plasma, EC-SOD A and EC-SOD B seem to be quantitatively important, whereas only minor amounts seem to be present in tissue. Studies up till now indicate that in the native environment, especially in the plasma, the three types of EC-SOD are present. In addition, EC-SOD forms an equilibrium  
30 between the plasma and endothelial surfaces.

Recombinant EC-SOD has previously been produced in mammalian cells, yeast and bacteria. Cost effective large scale production of recombinant EC-SOD has, however, not yet been possible.

- 5 Expression of recombinant genes in transgenic animals and preparation of transgenic animals in general have previously been reported. For example, U.S. Patent No. 4,736,866 discloses a transgenic mouse containing a c-myc oncogene. Oury et al. [54] report on the generation of a transgenic mouse
- 10 containing the EC-SOD cDNA under the control of the human  $\beta$ -actin promoter. The transgenic animals in these two disclosures were generated to be used as an animal disease model or for the study of the physiological function of a specific enzyme.
- 15 Reports of production of recombinant proteins in transgenic animals include PCT Publication No. WO 82/04443 (rabbit  $\beta$ -globin gene DNA fragment injected into the pronucleus of a mouse zygote); EPO Publication No. 0 264 166 (Hepatitis B surface antigen and Tissue Plasminogen Activator genes under
- 20 control of the whey acid protein promoter for mammary tissue specific expression); EPO Publication No. 0 247 494 (transgenic mice containing heterologous DNA encoding various forms of insulin); PCT Publication No. WO 88/00239 (tissue specific expression of DNA encoding factor IX under control of a whey
- 25 protein promoter); PCT Publication No. WO 88/01648 (transgenic mammal having mammary secretory cells incorporating a recombinant expression system comprising a mammary lactogen-inducible regulatory region and a structural region encoding a heterologous protein); EPO Publication No. 0 279 582 (tissue
- 30 specific expression of chloramphenicol acetyltransferase under control of rat  $\beta$ -casein promoter in transgenic mice); WO91/03551 (production of growth hormone in transgenic animal milk) and WO 91/08216 (production of recombinant polypeptides by bovine species and transgenic methods).

Genetic control elements from different milk protein genes have been used to direct production of recombinant proteins in the milk of transgenic animals [34-36]. The use of such transgenic techniques in farm animals offers the potential to  
5 produce pharmaceutical proteins in the milk.

Despite the fact that many cDNAs and genomic fragments now have been evaluated in mammary gland expression systems in transgenic animals, it is hard to define the factors that affect the level of expression. Limited attention has been  
10 focused on the qualitative aspects related to detailed analysis of post-translational modifications of recombinant proteins produced by this route; and production of an enzymatic functional multimeric metalloprotein in milk from a transgenic animal, where the function of the protein is closely  
15 related to the posttranslational modification, has not been described.

#### BRIEF DESCRIPTION OF THE INVENTION

It is an object of the invention to provide a means for producing recombinant human EC-SOD in a high yield.

20 Accordingly, in one aspect the present invention relates to a mammalian expression system comprising a DNA sequence encoding a polypeptide having the amino acid sequence shown in SEQ ID NO:2 or a variant thereof which has a biological activity of human EC-SOD said DNA sequence is combined with  
25 regulatory element of a gene encoding a milk protein of a mammal so as to form a hybrid gene which is expressible in the mammary gland of an adult female of a non-human mammal harbouring said hybrid gene so that the polypeptide encoded by the DNA sequence is produced when the hybrid gene is  
30 expressed.

In another aspect the present invention relates to a DNA fragment comprising a DNA sequence encoding a polypeptide comprising the amino acid sequence shown in SEQ ID NO:2 and

further comprising at least one intron sequence or a variant of said DNA sequence which

1) hybridizes with the DNA sequence shown in SEQ ID NO:1 or a specific part thereof, preferably under the stringent hybridization conditions or

2) encodes a polypeptide, the amino acid sequence of which is at least 85% homologous with the amino acid sequence shown in in SEQ ID NO:2, or

3) constitutes an effective subsequence of said DNA sequence,

and encodes a polypeptide having the biological activity of human EC-SOD.

The stringent hybridization conditions referred to above are to be understood in their conventional meaning, i.e. that hybridization is carried out at 67°C in 2xSSC and final washing at 67°C in 1xSSC using the method specified in the "Definition" part of the Examples below.

The term "homologous" is used here to illustrate the degree of identity between the amino acid sequence of a given polypeptide and the amino acid sequence shown in in SEQ ID NO:2. The amino acid sequence to be compared with the amino acid sequence shown in SEQ ID NO:2 may be deduced from a DNA sequence, e.g. obtained by hybridization as defined above, or may be obtained by conventional amino acid sequencing methods. The degree of homology is preferably determined on the amino acid sequence of a mature polypeptide, i.e. without taking any leader sequence into consideration. It is preferred that the degree of homology is at least 85%, such as at least 90%, preferably at least 95% or even 98% with the amino acid sequence shown in SEQ ID NO:2.

The term "effective subsequence" as used above refers to a subsequence which encodes a peptide being functional with respect to the activity of human EC-SOD as defined in the following. The subsequence may be the result of a truncation  
5 at either end of the DNA sequence or of the removal of one or more nucleotides or nucleotide sequences within DNA sequence.

The term "a biological activity" of human EC-SOD should be understood as the superoxide dismutating activity of EC-SOD.

The peptide encoded by the subsequence may have heparin  
10 affinity which is similar to, increased or decreased as compared to the heparin affinity of human EC-SOD type C.

In the present context, the term "heparin affinity which is similar to, increased or decreased as compared to the heparin affinity of human EC-SOD type C" indicates the polypeptide  
15 either has the same, or a less strong or stronger binding to heparin under physiological conditions than the binding of recombinant EC-SOD type C, the binding being assessed in vitro by elution with NaCl as described in the Examples below. Thus, the heparin affinity is determined by means of  
20 the concentration of NaCl required for eluting the polypeptide when bound to heparin.

In yet a further aspect, the present invention relates to a method of producing a transgenic non-human mammal capable of expressing a recombinant polypeptide of the invention, comprising injecting a mammalian expression system as defined  
25 above into a fertilized egg or a cell of an embryo of a mammal so as to incorporate the expression system into the germline of the mammal and developing the resulting injected fertilized egg or embryo into an adult female mammal.

30 A transgenic cell or animal contains one or more transgenes within its genome. A transgene is a DNA sequence integrated at a locus of a genome, wherein the transgenic DNA sequence is not otherwise normally found at that locus in that genome.

Transgenes may be made up of heterologous DNA sequences (sequences normally found in the genome of other species) or homologous DNA sequences (sequences derived from the genome of the same species).

- 5 As used herein, a "recombinant polypeptide" (or the recombinant DNA sequence encoding the same) is a "heterologous polypeptide". Heterologous polypeptides are polypeptides which are not normally produced by the transgenic animal.

- Each of the heterologous or homologous polypeptides are  
10 characterized by specific amino acid and nucleic acid sequences. It is to be understood, however, that such sequences include naturally occurring allelic variations thereof and variants produced by recombinant methods wherein such nucleic acid and polypeptide sequences have been modified by substitution, insertion and/or deletion of one or more nucleotides  
15 in said nucleic acid sequences to cause the substitution, insertion or deletion of one or more amino acid residues in the recombinant polypeptide. When the term DNA is used in the following, it should be understood that for the number of  
20 purposes where DNA can be substituted with RNA, the term DNA should be read to include RNA embodiments which will be apparent for the man skilled in the art.

- In further aspects, the present invention relates to a replicable expression vector which carries and is capable of  
25 expressing such DNA sequence, a cell harbouring such a vector, a method for producing the polypeptide, a transgenic animal *per se*, milk from such a transgenic animal.

#### DETAILED DESCRIPTION OF THE PRESENT INVENTION

- The mammalian expression system according to the invention  
30 may be an expression system comprising a DNA sequence encoding a polypeptide having the amino acid sequence shown in SEQ ID NO:2 or a variant thereof which has a biological activity of human EC-SOD, said DNA sequence is combined with



regulatory element of a gene encoding a milk protein of a mammal so as to form a hybrid gene which is expressible in the mammary gland of an adult female of a non-human mammal harbouring said hybrid gene so that the polypeptide encoded  
5 by the DNA sequence is produced when the hybrid gene is expressed.

As discussed in detail in the following, the expression system according to the invention is, for many purposes, preferably an expression system in which the DNA sequence  
10 contains at least one intron sequence, and, preferably, contains at least one permissive RNA splice signal.

As an example, the gene encoding the milk protein may be one selected from whey acidic protein (WAP) genes,  $\beta$ -lactoglobulin genes or casein genes. The present invention also comprises the hybrid gene as such.  
15

As mentioned above, the expression system is preferably one wherein the variant of the polypeptide encoded is at least 85% homologous with the amino acid sequence shown in SEQ ID NO:2. Another way of expressing the close structural relationship with the DNA sequence shown in SEQ ID NO:1 is to refer to hybridization: The expression system is preferably such that the DNA sequence encoding the polypeptide is one which hybridizes with the DNA sequence in SEQ ID NO:1 or a part thereof, preferably under stringent hybridization conditions as described in the examples.  
20  
25

An interesting embodiment comprises a modified DNA sequence which differs from the DNA sequence defined above in that at least one nucleotide has been deleted, substituted or modified or at least one additional nucleotide has been inserted so as to result in a DNA sequence which encodes a polypeptide having a biological activity of human EC-SOD.  
30

In another aspect, the present invention relates to a DNA fragment comprising a DNA sequence encoding a polypeptide having the amino acid sequence shown in SEQ ID NO:2 or variant thereof which has a biological activity of human EC-SOD, said DNA sequence comprising at least one intron sequence.

The DNA sequence is preferably one which

- 1) hybridizes with the DNA sequence shown in SEQ ID NO:1 or a specific part thereof, preferably under stringent hybridization conditions or
- 2) encodes a polypeptide, the amino acid sequence of which is at least 85% homologous with the amino acid sequence shown in SEQ ID NO:2, or
- 3) constitutes an effective subsequence of said DNA sequence.

In another embodiment the DNA fragment contains at least one permissive RNA splice signal.

In a preferred embodiment, the DNA fragment comprises substantially the DNA sequence shown in SEQ ID NO:1.

Alternatively, the DNA sequence may be a modified DNA sequence which differs from a DNA sequence as defined above in that at least one nucleotide has been deleted, substituted or modified or at least one additional nucleotide has been inserted so as to result in a DNA sequence which encodes a polypeptide having heparin affinity which is similar to, increased or decreased as compared to the heparin affinity of human EC-SOD, such as a variant selected from the group consisting of variants T216, T215, T213, T209, SAT216, SRT216, SA219, SA220, SA211-213, SA216, SA216/218 and SA216/220. The C-terminal sequences of these variants, where the differences are, are shown in Fig. 1 and the sequence listing.

In the present context, the term "gene" is used to indicate a DNA sequence which is involved in producing a polypeptide chain and which includes regions preceding and following the coding region (5'-upstream and 3'-downstream sequences) as well as intervening sequences, the so-called introns, which are placed between individual coding segments (so-called exons) or in the 5'-upstream or 3'-downstream region. The 5'-upstream region comprises a regulatory sequence which controls the expression of the gene, typically a promoter. The 3'-downstream region comprises sequences which are involved in termination of transcription of the gene and optionally sequences responsible for polyadenylation of the transcript and the 3' untranslated region.

The above mentioned regulatory or expression regulation sequences in addition to controlling transcription also contribute to RNA stability and processing, at least to the extent they are also transcribed.

Such expression regulation sequences are chosen to produce tissue-specific or cell type-specific expression of the recombinant DNA. Once a tissue or cell type is chosen for expression, 5' and optional 3' expression regulation sequences are chosen. Generally, such expression regulation sequences are derived from genes that are expressed primarily in the tissue or cell type chosen. Preferably, the genes from which these expression regulation sequences are obtained are expressed substantially only in the tissue or cell type chosen, although secondary expression in other tissue and/or cell types is acceptable if expression of the recombinant DNA in the transgene in such tissue or cell type is not detrimental to the transgenic animal. Particularly preferred expression regulation sequences are those endogenous to the species of animal to be manipulated. However, expression regulation sequences from other species such as those from human genes may also be used. In some instances, the expression regulation sequences and the structural DNA sequences (either geno-

mic or cDNA) are from the same species, e.g. each from bovine species or from a human source. In such cases, the expression regulation sequence and the DNA sequence are homologous to each other. Alternatively, the expression regulation sequences and DNA sequences (either cDNA or genomic) are obtained from different species, e.g. an expression regulation sequence from bovine species and a DNA sequence from a human source. In such cases, the expression regulation and DNA sequence are heterologous to each other. The following defines expression regulation sequences from endogenous genes. Such definitions are also applicable to expression regulation sequences from non-endogenous, heterologous genes.

In general, the 5' expression regulation sequence includes the transcribed portion of the endogenous gene upstream from the translation initiation sequence (the 5' untranslated region or 5' UTR) and those flanking sequences upstream therefrom which comprise a functional promoter. As used herein, a "functional promoter" includes those necessary untranscribed DNA sequences which direct the binding of RNA polymerase to the endogenous gene to promote transcription. Such sequences typically comprise a TATA sequence or box located generally about 25 to 30 nucleotides from the transcription initiation site. The TATA box is also sometimes referred to as the proximal signal. In many instances, the promoter further comprises one or more distal signals located upstream from the proximal signal (TATA box) which are necessary to initiate transcription. Such promoter sequences are generally contained within the first 100 to 200 nucleotides located upstream from the transcription initiation site, but may extend up to 500 to 600 nucleotides or more from the transcription initiation site. Such sequences are either readily apparent to those skilled in the art or readily identifiable by standard methods. Such promoter sequences alone or in combination with the 5' untranslated region are referred to herein as "proximal 5' expression regulation sequences".

In addition to such proximal 5' expression regulation sequences, it is preferred that additional 5' flanking sequences (referred to herein as "distal 5' expression regulation sequences") also be included in the transgene. Such distal 5' expression regulation sequences are believed to contain one or more enhancer and/or other sequences which facilitate expression of the endogenous gene and as a consequence facilitate the expression of the structural DNA sequence operably linked to the distal and proximal 5' expression regulation sequences. These 5' expression regulation sequences regulate the spatial and temporal distribution of gene expression. The amount of distal 5' expression regulation sequences depends upon the endogenous gene from which the expression regulation sequences are derived. In general, however, such sequences comprise 5' flanking regions of approximately 1 kb, more preferably 16 kb and most preferably about 30 kb of 5' flanking sequence. The determination of the optimal amount of distal 5' expression regulation sequences used from any particular endogenous gene is readily determined by varying the amount of distal 5' expression regulation sequence to obtain maximal expression. In general, the distal 5' expression regulation sequence will not be so large as to extend into an adjacent gene and will not include DNA sequences which adversely effect the level of transgene expression.

In addition, it is preferred that 3' expression regulation sequences also be included to supplement tissue or cell-type specific expression. Such 3' expression regulation sequences include 3' proximal and 3' distal expression regulation sequences from an appropriate endogenous gene. The 3' proximal expression regulation sequences include transcribed but untranslated DNA positioned downstream from the translation stop signal in the recombinant DNA sequence (also referred to as the 3' untranslated region or 3' UTR). Such sequences generally terminate at a polyadenylation sequence (either from the endogenous gene or from other sources such as SV40) and sequences that may affect RNA stability. Generally, 3' UTR's comprise about 100 to 1000 nucleotides or more down-

stream from the translation stop signal in the gene from which the 3' regulation sequence is derived. Distal 3' expression regulation sequences include flanking DNA sequences downstream from the proximal 3' expression regulation sequence. Some of these distal sequences are transcribed, but do not form part of the mRNA while other sequences in this 3' distal expression regulation sequence are not transcribed at all. Such distal 3' expression regulation sequences are believed to contain enhancer and/or other sequences which enhance expression. Such sequences are believed to be necessary for efficient polyadenylation and contain transcription termination sequences. Preferably, such sequences comprise about 2 kb, more preferably 8 kb and most preferably about 15 kb of 3' flanking sequence.

Although the use of both 5' and 3' expression regulation sequences are preferred, in some embodiments of the invention, endogenous 3' regulation sequences are not used. In such cases, the 3' proximal expression regulation sequences normally associated with the genomic DNA encoded by the recombinant DNA sequence are used to direct polyadenylation. In addition, distal 3' regulation sequences from the genomic DNA encoding the recombinant polypeptide may also be employed preferably in the same amounts as set forth for endogenous 3' expression regulation sequences. In such cases, it is to be understood that the recombinant polypeptide encoded by the transgene may comprise either genomic DNA or a double stranded DNA derived from cDNA. As with the 5' expression regulation sequences, the optimal amount of 3' expression regulation sequence may be readily determined by varying the amount of 3' flanking sequence to obtain maximal expression of the recombinant polypeptide. In general, the distal 3' regulation sequence, be it from an endogenous gene or a heterologous gene, will not extend into the adjacent gene from which it is derived and will exclude any sequences which adversely effect the level of transgene expression.

In addition to the 5' and 3' expression regulation sequences and the recombinant DNA (either genomic or derived from cDNA) the transgenes of the invention may also comprise an intron sequence which interrupts the transcribed region of the trans-  
5 gene. Recombinant intervening sequences may, however, also comprise a "hybrid intervening sequence". Such hybrid intervening sequences comprise a 5' RNA splice signal and 3' RNA splice signal from intervening sequences from heterologous or homologous sources.

- 10 Such hybrid intervening sequences containing permissive RNA splice signals are preferably used when the recombinant DNA corresponds to a cDNA sequence.

Based on the foregoing, it is apparent that preferred transgenes include large amounts of 5' and 3' expression regula-  
15 tion sequences. Further, the recombinant DNA is preferably derived from genomic clones which may be tens to hundreds of kilobases in length. Based on the present technology for cloning and manipulating DNA, the construction and micro-injection of transgenes is practically limited to linearized  
20 DNA having a length not greater than about 50 kb. However, the transgenes of the invention, especially those having a length greater than about 50 kb, may be readily generated by introducing two or more overlapping fragments of the desired transgene into an embryonal target cell. When so introduced,  
25 the overlapping fragments undergo homologous recombination which results in integration of the fully reconstituted transgene in the genome of the target cell. In general, it is preferred that such overlapping transgene fragments have 100% homology in those regions which overlap. However, lower se-  
30 quence homology may be tolerated provided efficient homologous recombination occurs. If non-homology does exist between the homologous sequence portions, it is preferred that the non-homology not be spread throughout the homologous sequence portion but rather be located in discrete areas. Although as  
35 few as 14 base pairs at 100% homology are sufficient for homologous recombination in mammalian cells [55], longer homolo-

gous sequence portions are preferred, e.g. 500 bp, more preferably 1000 bp, next most preferably 2000 bp and most preferably greater than 2000 bp for each homologous sequence portion.

- 5 When the transgene of the invention encodes a recombinant polypeptide that is encoded by recombinant DNA derived from or corresponding to genomic DNA (or comprised substantially of such genomic sequences, e.g. greater than about 50%, more preferably greater than about 75%, most preferably greater
- 10 than 90% of the codons encoding the recombinant polypeptide are from genomic sequences), the molar concentrations and protein levels in transgenic milk are the same as for cDNA or higher. In general, the molar concentration of the recombinant polypeptide in such transgenic milk is preferably greater
- 15 er than about 50  $\mu$ M, more preferably greater than about 150  $\mu$ M, most preferably greater than about 500  $\mu$ M.

The foregoing molar concentration and protein levels in bovine transgenic milk will vary depending upon the molecular weight of the particular recombinant polypeptide. A parti-

20 cular advantage of producing a recombinant polypeptide in transgenic milk is that relatively large molecular weight polypeptides may be so produced which are otherwise difficult to produce in large quantities in other systems such as prokaryotic expression systems.

- 25 The mouse, however, normally produces between 55 to 80 milligrams of protein per ml of milk, and a rabbit produces about 100 milligrams of protein per ml of milk. A cow, on the other hand, normally produces between 30 to 34 milligrams of protein per ml. Since exceptionally high levels of recombinant
- 30 polypeptide production may adversely affect the production of endogenous milk protein and/or have adverse effects upon the mammary secretory gland, it is preferred that the recombinant polypeptide concentration be between about 1 and 50% of the normal milk protein concentration, more preferably between 10
- 35 to 20% and most preferably between 10 and 15% of the normal



amount of protein produced in bovine milk. Such preferred ranges also provide a preferred maximum limit to the aforementioned levels of protein produced in transgenic bovine milk.

- 5 The term "effective subsequence" of the gene is to be understood in the same manner as defined above in connection with the DNA sequence.

The hybridization may be carried out as described in the "Definition" part of the Examples below, preferably on the  
10 basis of a probe comprising the coding part of the DNA sequence shown in the SEQ ID NO:1 below. The terms "homologous" and "effective subsequences" are used in a similar manner as that defined above.

Preferably, the polypeptide encoded by the variant of the DNA  
15 sequence is at least 90% homologous, such as at least 95% or even 98% homologous with the amino acid sequence shown in SEQ ID NO:2.

Examples of specific variants of the DNA sequence of the invention are DNA sequences which comprises an essential part  
20 of or the complete DNA sequence shown in SEQ ID NO:1 particularly adapted for expression in a transgenic animal. This DNA sequence is one which, when inserted in the expression system together with suitable regulatory sequences, results in the expression of a polypeptide having the amino acid sequence  
25 shown in SEQ ID NO:2 or a variant or a subsequence thereof.

As mentioned above, the DNA sequence shown in SEQ ID NO:1 encodes a polypeptide comprising the functional domain/domains of human EC-SOD. While the presence of a signal peptide in most cases is a prerequisite for allowing the polypeptide  
30 expressed from the DNA sequence to be transported out of the cell in which it is produced, the nature and origin of the particular signal peptide to be used may vary and need not be

the signal peptide naturally associated with the human EC-SOD.

Native human EC-SOD is glycosylated at asparagine at position 89. It is contemplated that an EC-SOD variant comprising more  
5 residues which are glycosylated than the native EC-SOD will have a higher heparin affinity than the native EC-SOD.

The glycosylation of a recombinant polypeptide is dependent of the selected expression system. It is well known that eukaryotic cells of different species and/or tissue origin  
10 show variation in the glycosylation machinery. Thus, to achieve the glycosylation modifications of interest, it is critical to select a host organism for the production of the recombinant molecule, which have the capacity to perform the appropriate post-translational glycosylation modifications.

15 However, there are methods available that allow the modification of the glycosylation machinery of a host organism. This can be done by altering the genome of the host organism, for example a host cell or a transgenic animal, by introduction of recombinant genetic elements. These genetic elements can  
20 either encode additional or modified glycosyltransferases or other involved enzymes, and mediate their expression, or inhibit the function of endogenous glycosyltransferases or other involved enzymes. Inhibition can be achieved by knocking-out endogenous glycosyltransferase gene functions or by  
25 introduction of vectors encoding RNA sequences which are complementary to endogenous glycosyltransferase mRNA species, thereby function as antisense RNA.

The polypeptide encoded by the modified DNA sequence has normally an amino acid sequence which is different from the  
30 amino acid sequence of the human EC-SOD.

When "substitution" is performed, one or more nucleotides in the full nucleotide sequence are replaced with one or more different nucleotides; when "addition" is performed, one or

more nucleotides are added at either end of the full nucleotide sequence; when "insertion" is performed one or more nucleotides within the full nucleotide sequence is inserted; and when "deletion" is performed one or more nucleotides are  
5 deleted from the full nucleotide sequence whether at either end of the sequence or at any suitable point within it.

A modified DNA sequence may be obtained by well-known methods, e.g., by use of site-directed mutagenesis.

An example of an important modified DNA sequence of the  
10 invention is a DNA sequence in which additional codons encoding asparagine residues have been inserted so as to result in a modified DNA sequence encoding a polypeptide having an increased number of residues to be glycosylated. The additional residues may be inserted either by being added at either  
15 end or within a DNA sequence of the invention or by replacing one or more non-asparagine codons present in a DNA sequence of the invention. A polypeptide encoded by such a modified DNA sequence is contemplated to have a higher degree of glycosylation.

20 Another example of interesting modified DNA sequences are DNA sequences encoding variants of EC-SOD having a reduced heparin affinity as compared to recombinant EC-SOD type C. Such variants may be variants selected from the group consisting of variants T216, T215, T213, T209, SAT216, SRT216, SA219,  
25 SA220 and SA211-213 (see Fig. 1 and the sequence listing wherein sequences SEQ ID NOS: 8-15, 18-23 and 28-31 show the C-terminal sequences of these variants).

Further examples of interesting modified DNA sequences are DNA sequences encoding variants of EC-SOD having an increased  
30 heparin affinity as compared to recombinant EC-SOD type C. Such variants may be variants selected from the group consisting of variants SA216, SA216/218 and SA216/220. The C-terminal sequences of these variants are shown as SEQ ID NOS: 16-17, 24-25 and 26-27, respectively.

For the purpose of preparing modified DNA sequences, site-directed mutagenesis may be carried out using specific oligonucleotide probes conferring an exchange/removal of the relevant amino acids residues.

- 5 The DNA sequences used in the mammalian expression system of the invention explained herein may comprise natural as well as synthetic DNA sequences, the natural sequence typically being derived directly from cDNA or genomic DNA, normally of mammalian origin, e.g. as described below. A synthetic sequence may be prepared by conventional methods for synthetically preparing DNA molecules. Of course, also the DNA sequence may be of mixed cDNA and genomic, mixed cDNA and synthetic cDNA, and mixed genomic DNA and synthetic DNA origin. Also RNA sequences may be used as described above.
- 10
- 15 The terms "sequence", "subsequence", "variant" and "polypeptide" as used herein with respect to sequences, subsequences, variants and polypeptides according to the invention should of course be understood as not comprising these phenomena in their natural environment, but rather fragments, e.g., in isolated, purified, *in vitro* or recombinant form. When reference is made to a DNA sequence of the invention this should be understood to include "variants", "subsequences" and "modified sequences" as defined above. The term "subsequence" is to be understood as a part of a longer sequence, in the present context as a part of the sequences shown in e.g. SEQ ID NO:1 or 2. Similarly, when reference is made to "a polypeptide of the invention", this should be understood to include any of the polypeptides defined herein.
- 20
- 25

In a further aspect, the present invention relates to a replicable expression vector which carries and is capable of mediating the expression of a DNA sequence encoding human EC-SOD.

30

In the present context, the term "replicable" means that the vector is able to replicate in a given type of host cell into

which it has been introduced. Immediately upstream of the human EC-SOD DNA sequence there may be provided a sequence coding for a signal peptide, the presence of which ensures secretion of the human EC-SOD expressed by host cells harbouring the vector. The signal sequence may be the one naturally associated with the human EC-SOD DNA sequence or of another origin.

The vector may be any vector which may conveniently be subjected to recombinant DNA procedures, and the choice of vector will often depend on the host cell into which it is to be introduced. Thus, the vector may be an autonomously replicating vector, i.e. a vector which exists as an extrachromosomal entity, the replication of which is independent of chromosomal replication; examples of such a vector are a plasmid, phage, cosmid, mini-chromosome or virus. Alternatively, the vector may be one which, when introduced in a host cell, is integrated in the host cell genome and replicated together with the chromosome(s) into which it has been integrated. The vector of the invention may carry any of the DNA sequences of the invention as defined above and be used for the expression of any of the polypeptides of the invention defined above.

The present invention thus also relates to a replicable expression vector designated pS 172 which has been deposited on 7 June 1993 with the collection of Deutsche Sammlung von Mikroorganismen und Zellkulturen GmbH (DSMZ) under the accession number DSM 8335 in accordance with the provisions of the Budapest Treaty, and expression vectors expressing DNA sequences which differ from the DNA sequences of the said deposited expression vector, but which code for the same polypeptide or a variant thereof which has a biological activity of human EC-SOD as well as a replicable expression as defined above, wherein the DNA sequence expressed is one which differs from the DNA sequence of the deposited vector in that at least one nucleotide has been deleted, substituted or modified or at least one additional nucleotide has been

inserted so as to result in a DNA sequence which encodes a polypeptide having a biological activity of EC-SOD.

The present invention further relates to a cell harboring a replicable expression vector as defined above. In principle, this cell may be of any type of cell, i.e. a prokaryotic cell such as a bacterium, e.g. *E. coli*, a unicellular eukaryotic organism, a fungus or yeast, e.g. *Saccharomyces cerevisiae* or a cell derived from a multicellular organism, e.g. a mammal. The mammalian cells are especially suitable for the purpose and are further discussed below.

In another important aspect, the invention relates to a method of producing recombinant human EC-SOD, said method comprising introducing at least one expression system as defined above into the genome of a non-human mammal in such a way that the DNA encoding the polypeptide is expressed in a mammary gland of the non-human mammal, collecting the milk secreted from the gland, and recovering the recombinant polypeptide from the milk, optionally followed by purification. A suitable vector may be any of the vectors described above, and an appropriate host cell may be any of the cell types listed above. The methods employed to construct the vector and affect introduction thereof into the host cell may be any methods known for such purposes within the field of recombinant DNA.

In one embodiment 1-10 copies of the expression system or a subsequence thereof are injected, in another 1-7 copies of the expression system or a subsequence thereof are injected.

The copies of the expression system or of a subsequence thereof may either be identical or comprise different DNA sequences resulting in expression of at least two different variants of EC-SOD, resulting in heterologous tetramers of EC-SOD in the milk. The variants may have the same heparin affinity or differ with respect to heparin affinity.

In order to be secreted, the DNA sequence encoding human EC-SOD should be preceded by a sequence encoding a signal peptide, the presence of which ensures secretion of human EC-SOD from the cells so that at least a significant proportion of the human EC-SOD expressed is secreted, and may be recovered.

The mammary gland as a tissue of expression and genes encoding milk proteins are generally considered to be particularly suitable for use in the production of heterologous proteins in transgenic non-human mammals as milk proteins are naturally produced at high expression levels in the mammary gland. Also, milk is readily collected and available in large quantities.

In the present context the term "hybrid gene" denotes a DNA sequence comprising on the one hand a DNA sequence encoding human EC-SOD as defined above and on the other hand a DNA sequence of the milk protein gene which is capable of mediating the expression of the hybrid gene product. The term "gene encoding a milk protein" or "milk protein gene" denotes an entire gene as well as an effective subsequence thereof capable of mediating and targeting the expression of the hybrid gene to the tissue of interest, i.e. the mammary gland. The milk protein gene may be the gene for  $\beta$ -lactoglobulin,  $\alpha$ -lactalbumin or a casein, but the whey acid protein gene is particularly preferred. Normally, the effective subsequence is one which at least harbours one or more of a promoter region, a transcriptional start site, 3' and 5' non-coding regions and structural sequences. The DNA sequence encoding human EC-SOD is preferably substantially free from prokaryotic sequences, such as vector sequences, which may be associated with the DNA sequence after, e.g., cloning thereof.

The hybrid gene is preferably formed by inserting *in vitro* the DNA sequence encoding human EC-SOD into the milk protein gene by use of techniques known in the art. Alternatively,

the DNA sequence encoding human EC-SOD can be inserted in vivo by homologous recombination.

- Normally, the DNA sequence encoding human EC-SOD will be inserted in one of the first exons of the milk protein gene of choice or an effective subsequence thereof comprising the first exons and preferably a substantial part of the 5' flanking sequence which is believed to be of regulatory importance.
- 10 The hybrid gene preferably comprises a sequence encoding a signal peptide so as to enable the hybrid gene product to be secreted correctly into the mammary gland. The signal peptide will typically be the one normally found in the milk protein gene in question or one associated with the DNA sequence
- 15 encoding human EC-SOD. However, also other signal sequences capable of mediating the secretion of the hybrid gene product to the mammary gland are relevant. Of course, the various elements of the hybrid gene should be fused in such a manner as to allow for correct expression and processing of the gene
- 20 product. Thus, normally the DNA sequence encoding the signal peptide of choice should be precisely fused to the N-terminal part of the DNA sequence encoding human EC-SOD. In the hybrid gene, the DNA sequence encoding human EC-SOD will normally comprise its stop codon, but not its own message cleavage
- 25 and polyadenylation site. Downstream of the DNA sequence encoding human EC-SOD, the mRNA processing sequences of the milk protein gene will normally be retained.

- A number of factors are contemplated to be responsible for the actual expression level of a particular hybrid gene. The
- 30 capability of the promoter as well of other regulatory sequences as mentioned above, the integration site of the expression system in the genome of the mammal, the integration site of the DNA sequence encoding human EC-SOD in the milk protein encoding gene, elements conferring post-transcriptional regulation and other similar factors may be of
- 35 vital importance for the expression level obtained. On the



basis of the knowledge of the various factors influencing the expression level of the hybrid gene, the person skilled in the art would know how to design an expression system useful for the present purpose.

- 5 The milk protein gene to be used may be derived from the same species as the one in which the expression system is to be inserted, or it may be derived from another species. In this connection it has been shown that the regulatory elements that target gene expression to the mammary gland are functional across species boundaries (which may be due to a possible common ancestor) [35].

Examples of suitable genes encoding a milk protein or effective subsequences thereof to be used in the construction of an expression system of the invention are normally found among whey proteins of various mammalian origins, e.g. a whey acidic protein (WAP) gene, preferably of murine origin, and a  $\beta$ -lactoglobulin gene, preferably of ovine origin. Also casein genes of various origins may be found to be suitable for the transgenic production of human EC-SOD, e.g. bovine aS1-casein and rabbit  $\beta$ -casein. The presently preferred gene is a murine WAP gene as this has been found to be capable of providing a high level expression of a number of foreign human proteins in milk of different transgenic animals [35].

Another sequence preferably associated with the expression system of the invention is a so-called expression stabilizing sequence capable of mediating high-level expression. Strong indications exist that such stabilizing sequences are found in the vicinity of and upstream of milk protein genes.

The DNA sequence encoding a human EC-SOD to be inserted in the expression system of the invention may be of cDNA, genomic or synthetic origin or any combination thereof. While some expression systems have been found to function best when cDNA encoding a desirable protein is used, others have been found to require the presence of introns and other regulatory

regions in order to obtain a satisfactory expression [35]. In some cases it may be advantageous to introduce genomic structures in vector constructs compared to cDNA elements [56]. The intron and exon structure may result in higher steady state mRNA levels than obtained when cDNA based vectors are used.

In the specification, the term "intron" includes the whole of any natural intron or part thereof.

In a further aspect, the present invention relates to a hybrid gene comprising a DNA sequence encoding human EC-SOD inserted into a gene encoding a milk protein of a mammal, the DNA sequence being inserted in the milk protein gene in such a manner that it is expressible in the mammary gland of an adult female of a non-human mammal harboring the hybrid gene. The hybrid gene and its constituents have been discussed in detail above. The hybrid gene constitutes an important intermediate in the construction of an expression system of the invention as disclosed above.

In another aspect, the present invention relates to a non-human mammalian cell harboring an expression system as defined above. The mammalian cell is preferably an embryo cell or a pro-nucleus. The expression system is suitably inserted in the mammalian cell using a method as explained in the following.

In a further important aspect, the present invention relates to a method of producing a transgenic non-human mammal capable of expressing human EC-SOD, comprising injecting an expression system of the invention as defined above into a fertilized egg or a cell of an embryo of a mammal so as to incorporate the expression system into the germline of the mammal and developing the resulting injected fertilized egg or embryo into an adult female mammal.

The "non-human mammals" of the invention comprise all non-human mammals capable of producing a "transgenic non-human mammal" having a "desirable phenotype". Such mammals include rabbits, sheep, goats, murine species, bovine species, canine species, etc.

Desirable phenotypes for transgenic non-human mammals include, but are not limited to, the production of recombinant polypeptides in the milk of female transgenic non-human mammals.

10 The transgenic non-human mammals of the invention are produced by introducing a "transgene" into an embryonal target cell of the animal of choice. In one aspect of the invention, a transgene is a DNA sequence which is capable of producing a desirable phenotype when contained in the genome of cells of  
15 a transgenic non-human mammal. In specific embodiments, the transgene comprises a "recombinant DNA sequence" encoding a "recombinant polypeptide". In such cases, the transgene is capable of being expressed to produce the recombinant polypeptide.

20 The incorporation of the expression system into the germline of the mammal may be performed using any suitable technique, e.g. as described in [41] or in WO 91/08216.

Methods of introducing transgenes or overlapping transgene fragments into embryonal target cells include microinjection  
25 of the transgene into the pronuclei of fertilized oocytes or nuclei of ES cells of the non-human animal. Such methods for murine species are well known to those skilled in the art. Alternatively, the transgene may be introduced into an animal by infection of zygotes with a retrovirus containing the  
30 transgene [57]. The preferred method is microinjection of the fertilized oocyte. In this preferred embodiment, the fertilized oocytes are first microinjected by standard techniques. They are thereafter cultured *in vitro* until a "pre-implantation embryo" is obtained. Such pre-implantation embryos pre-

ferably contain approximately 16 to 150 cells. The 16 to 32 cell stage of an embryo is commonly referred to as a morula. Those pre-implantation embryos containing more than 32 cells are commonly referred to as blastocysts. They are generally characterized as demonstrating the development of a blastocoel cavity typically at the 64 cell stage. Methods for culturing fertilized oocytes to the pre-implantation stage include those described by Gordon et al. [58]; Hogan et al. [41] (for the mouse embryo); and Hammer et al. [59] (for rabbit and porcine embryos); Gandolfi et al. [60]; Rexroad et al. [61] (for ovine embryos); and Eyestone et al. [62]; Camous et al. [63]; and Heyman et al. [64] (for bovine embryos). Such pre-implantation embryos are thereafter transferred to an appropriate female by standard methods to permit the birth of a transgenic or chimeric animal depending upon the stage of development when the transgene is introduced. As is well known, mosaic animals can be bred to form true germ-line transgenic animals.

Since the frequency of transgene incorporation is often low, the detection of transgene integration in the pre-implantation embryo is highly desirable. In one aspect of the invention, methods are provided for identifying embryos wherein transgenesis has occurred and which permit implantation of transgenic embryos to form transgenic animals. In this method, one or more cells are removed from the pre-implantation embryo. When equal division is used, the embryo is preferably not cultivated past the morula stage (32 cells). Division of the pre-implantation embryo (reviewed by Williams et al. [65]) results in two "hemi-embryos" (hemi-morula or hemi-blastocyst) one of which is capable of subsequent development after implantation into the appropriate female to develop in utero to term. Although equal division of the pre-implantation embryo is preferred, it is to be understood that such an embryo may be unequally divided either intentionally or unintentionally into two hemi-embryos which are not necessarily of equal cell number. Essentially, all that is required is that one of the embryos which is not analyzed as

hereinafter described be of sufficient cell number to develop to full term in utero. In a specific embodiment, the hemi-embryo which is not analyzed as described herein, if shown to be transgenic, is used to generate a clonal population of  
5 transgenic non-human animals.

One of each of the hemi-embryos formed by division of pre-implantation embryos is analyzed to determine if the transgene has been integrated into the genome of the organism. Each of the other hemi-embryos is maintained for subsequent  
10 implantation into a recipient female of the species.

The early identification of the pre-implantation embryos containing the integrated transgene is achieved by analyzing the DNA from one of each of the hemi-embryos. Such DNA is typically obtained by lysing the hemi-embryo and analyzing  
15 the thus released DNA. A polymerase chain reaction is performed to amplify all or part of the transgene. When the entire transgene is amplified, two extension primers each complementary to opposite strands at opposing ends of the transgene are used for amplification. Generally, the amplified DNA from  
20 the hemi-embryo is subjected to electrophoresis followed by hybridization with labeled probe complementary to the region of the transgene between the two extension primers. This facilitates the determination of the size of the amplified DNA sequences, if any, and provides an indication of whether  
25 the transgene has been integrated into the pre-implantation embryo from which the hemi-embryo was obtained (now called a "transgenic hemi-embryo"). If it has, the remaining untreated transgenic hemi-embryo is transplanted into a recipient  
parent. After in utero development, the transgenic non-human  
30 animal having the desired phenotype conferred by the integrated transgene is identified by an appropriate method in utero or after birth.

The above described methods for the detection of transgenesis in pre-implantation embryos provide economical and time  
35 saving methods for generating transgenic non-human animals

since they significantly decrease the number of pregnancies required to produce a transgenic animal and substantially increase the likelihood that an implanted embryo will produce a transgenic non-human animal. Such methods are especially  
5 important for those animals for which very low or non-existent frequencies of transgenesis have been obtained, e.g. bovine species.

In an alternate embodiment, the above described method for detecting transgenesis in pre-implantation embryos is combined with embryonic cloning steps to generate a clonal population of transgenic embryos which may thereafter be implanted into recipient females to produce a clonal population of transgenic non-human animals also having the same genotype. In this regard, it is to be understood that transgenic embryos and/or non-human transgenic animals having the same  
15 "genotype" means that the genomic DNA is substantially identical between the individuals of the embryo and/or transgenic animal population. It is to be understood, however, that during mitosis various somatic mutations may occur which may produce variations in the genotype of one or more cells  
20 and/or animals. Thus, a population having the same genotype may demonstrate individual or subpopulation variations.

After a hemi-embryo is identified as a transgenic hemi-embryo, it is cloned. Such embryo cloning may be performed by  
25 several different approaches. In one cloning method, the transgenic hemi-embryo is cultured in the same or in a similar medium as used to culture individual oocytes to the pre-implantation stage. The "transgenic embryo" so formed (preferably a transgenic morula) is then divided into "transgenic  
30 hemi-embryos" which can then be implanted into a recipient female to form a clonal population of two transgenic non-human animals. Alternatively, the two transgenic hemi-embryos obtained may be again cultivated to the pre-implantation stage, divided, and recultivated to the transgenic embryo  
35 stage. This procedure is repeated until the desired number of clonal transgenic embryos having the same genotype are obtained.

ned. Such transgenic embryos may then be implanted into recipient females to produce a clonal population of transgenic non-human animals.

In a preferred cloning method, the transgenic embryo is  
5 cloned by nuclear transfer according to the techniques of Prather et al. [66]; Roble et al. [67]. According to this method, nuclei of the transgenic embryo are transplanted into enucleated oocytes, each of which is thereafter cultured to the blastocyst stage. At this point, the transgenic embryos  
10 may be resubjected to another round of cloning by nuclear transplantation or may be transferred to a recipient parent for production of transgenic offspring having the same genotype.

In addition to the foregoing methods for detecting early  
15 transgenesis, other methods may be used to detect transgenesis. Such method include *in utero* and *post partum* analysis of tissue. *in utero* analysis is performed by several techniques. In one, transvaginal puncture of the amniotic cavity is performed under echoscopic guidance [68, 69]. This involves  
20 recovering about 15 to 20 milliliters of amniotic fluid between about day 35 and day 100 of gestation. This volume of amniotic fluid contains about 1000 to 12,000 cells per ml originating from the urogenital tract, the skin and possibly the lungs of the developing embryo. Most of these cells are  
25 dead. Such cells, however, contain genomic DNA which is subjected to PCR analysis for the transgene as an indication of a successful transgenesis. Alternatively, fetal cells may be recovered by chorion puncture. This method may also be performed transvaginally and under echoscopic guidance. In  
30 this method, a needle is used to puncture the recipient animal's placenta, particularly the placentonal structures, which are fixed against the vaginal wall. Such sampling may be performed around day 60 of gestation in bovine species. Chorion cells, if necessary, are separated from maternal  
35 tissue and subjected to PCR analysis for the transgene as an indication of successful transgenesis.

Transgenesis may also be detected after birth. In such cases, transgene integration can be detected by taking an appropriate tissue biopsy such as from the ear or tail of the putative transgenic animal. About one to two centimeters of tail or  
5 about five to ten square millimeters of ear are obtained followed by southern blotting with a probe for the transgene according to the method of [42].

Normally, not all of the injected eggs will develop into transgenic mammals capable of expressing human EC-SOD. Trans-  
10 genic founder animals can be identified e.g. as described in Example 2. About half of the mammals will from a statistical point of view be males. On the basis of the identified transgenic individuals - male and female - progeny can be established and stable lines of transgenic animals established.

15 Once integrated in the germ line, the DNA sequence encoding human EC-SOD may be expressed at high levels to produce a correctly processed and functional human EC-SOD. Transgenic females from which recombinant polypeptide can be harvested can thus be bred in the following generations.

20 In a further aspect, the present invention relates to a transgenic non-human mammal prepared by a method as described above.

The DNA used to make transgenic cells and animals preferably comprises genomic DNA rather than cDNA. This is because the  
25 expression of transgenes is preferably limited to tissue-specific expression as well as temporal-specific expression. When the transgene is derived from genomic DNA, important cis-acting regulatory sequences such as enhancers and other regulatory elements, located either in introns or in regions  
30 distant from the structural gene, can be included. Such regulatory sequences are lost during transcription and RNA processing and accordingly are not generally available with cDNA-derived transgenes.



In a further aspect, the present invention relates to a transgenic non-human mammal prepared by a method as described above.

While the transgenic non-human mammal of the invention in its broadest aspect is not restricted to any particular type of mammal, the mammal will normally be selected from the group consisting of rabbits, mice, rats, sheep, pigs, goats, lamas, camels and cattle. For large scale production of human EC-SOD the larger animals such as sheep, goats, pigs and especially cattle are normally preferred due to their high milk production. However, also mice, rabbits and rats may be interesting due to the fact that the manipulation of these animals is more simple and results in transgenic animals more quickly than when, e.g. cattle, are concerned.

Also progeny of a transgenic mammal as defined above, capable of producing human EC-SOD is within the scope of the present invention.

From the above explanation it will be clear that the present invention for the first time makes it possible to produce milk from a non-human mammal comprising human EC-SOD, the importance and utility of which will be apparent from the present context. Thus, in a further aspects of the present invention includes milk from a non-human mammal comprising recombinant human EC-SOD. Of particular interest is milk from a non-human mammal comprising a polypeptide of the invention as defined above comprising the amino acid sequence shown in SEQ ID NO:2 or a polypeptide encoded by a DNA sequence or a variant or subsequence thereof as defined above. Typically, the milk of the invention will be obtained from a transgenic mammal of the invention as defined above.

In another aspect, the present invention relates to a method of obtaining human EC-SOD comprising collecting milk from a transgenic non-human mammal of the invention as defined above and recovering the human EC-SOD from the milk. The milk may

be collected in any suitable manner normally used in connection with the collection of milk from the mammal in question.

The polypeptide of the invention having the superoxide dismutating property of native EC-SOD may be used for the diagnosis, prophylaxis or treatment of diseases or disorders connected with the presence or formation of superoxide radicals and other toxic oxygen intermediates derived from the superoxide radical.

Examples of such diseases or disorders are selected from conditions involving ischaemia followed by reperfusion, e.g. infarctions such as heart, kidney, brain, spinal cord, or intestinal infarctions; transplantation of organs such as heart, lung, pancreas, liver, skin, bone tissue, severed extremities, skeletal muscle; inflammatory diseases such as rheumatoid arthritis, pancreatitis, in particular acute pancreatitis, pyelonephritis and other types of nephritis, and hepatitis, neuritis, uveitis, cystitis, peyronies disease, autoimmune diseases, insulin-dependent diabetes mellitus, disseminated intravascular coagulation, fatty embolism, haemorrhagic shock, endotoxin-induced shock, septicemia, severe viral infections, adult respiratory distress, infantile respiratory distress, brain haemorrhages in neonates, burns, preservation of lens and cornea in the context of transplantation, adverse effects of ionizing radiation, carcinogenesis, and adverse effects of toxins such as alloxan, paraquat, and some cytostatic compounds.

Thus, the polypeptide of the invention is indicated for substantially the same applications as CuZn SOD the therapeutic activity of which has been more thoroughly documented as discussed below. EC-SOD and the EC-SOD variants of the invention have, however, been found to possess a number of characteristics which are assumed to make them particularly useful for therapeutic applications.

CuZn SOD has a low molecular weight (33,000) which causes it to become eliminated very quickly by glomerular filtration in the kidneys so that in rodents it has a plasma half-life of less than 10 minutes and, in human beings, the enzyme has a half-life of about 20-30 minutes. EC-SOD C [15] and the EC-SOD variants have a much longer half-life. This is partly due to the high molecular weight of EC-SOD which prevents it from being eliminated by glomerular filtration, and partly due to the fact that EC-SOD and EC-SOD variants having heparin affinity of the C-type seem to bind to endothelial cell surfaces. In the therapeutic use of EC-SOD and variants according to the invention, the enzyme therefore preferably has a half-life in human beings of at least 4 hours and possibly even longer.

As explained above, EC-SOD is, in its native environment, a secreted protein and it is therefore likely that it is specifically synthesized for a function in extracellular space (in extracellular fluids or on cell surfaces) which may cause it to exhibit properties which are particularly well adapted to protect plasma components or the outer surface of cells against the toxic effects of superoxide radicals or other oxygen radicals. A fundamental property of EC-SOD C, and most likely also a polypeptide having a slightly reduced affinity for heparin compared to the heparin affinity of recombinant EC-SOD C, e.g. the variant T216, is its affinity for heparan sulfate which *in vivo* apparently has the correlate of binding to heparan sulfate proteoglycan in the glycocalyx of cell surfaces and in the interstitial connective tissue matrix. This property would seem to be a particularly efficient way to protect cells and tissues against an external superoxide radical source.

The significance of the apparent association of EC-SOD and EC-SOD variants with cell membranes is further supported by the finding that CuZn SOD which has been modified with polylysine to bind to cell membranes is better able to protect activated (superoxide radical-producing) polymorphonuclear

leukocytes (PMN) from autoinactivation (cell death) than normal CuZn SOD which is negatively charged and therefore tends to be repelled by the cell membranes. The fact that *Nocardia asteroides* possesses a membrane-associated SOD which appears to confer efficient protection against activated human PMNs as the susceptibility of *Nocardia* to PMNs is significantly increased when *Nocardia* cells are treated with antibodies towards this SOD also points to a cell membrane-protective function of SOD bound to cell surfaces. Unlike EC-SOD and EC-SOD variants, CuZn SOD has an intracellular function which may make it less well suited for extracellular application, i.e. occasioned by the extracellular presence of superoxide radicals. Furthermore, its brief half-life compared to that of EC-SOD and EC-SOD variants mentioned above would seem to make it necessary to administer larger doses at shorter intervals than is likely to be the case with the EC-SOD variants of the invention.

Parenterally administered CuZn SOD has been shown to exhibit an anti-inflammatory effect in a series of animal models of inflammation as well as in inflammatory diseases in animals. In humans, positive effects of CuZn SOD have been reported in rheumatoid arthritis and arthroses, in inflammation of the bladder and other urological conditions.

Parenterally administered CuZn SOD is not taken up by the cells and must exert its activity in the extracellular space. CuZn SOD encapsulated in liposomes is taken up by the cells and is reported to be effective against Crohn's disease, Bechet's disease, dermatitis herpetiformis, ulcerative colitis, Kawasaki's disease and against the adverse effects of radiation therapy. The mechanism of the anti-inflammatory activity of CuZn SOD is not quite clear. Direct protection against oxygen radicals formed by activated leukocytes has been suggested. Another possibility is prevention of the formation of a superoxide induced strongly chemotaxic substance.

The other large potential area of application for SOD is as a protective factor against tissue damage caused by ischaemia followed by reperfusion. If the supply of blood to a tissue is cut off, the tissue will slowly become necrotic. Macro- and microscopically the damage will typically develop slowly over many hours. If the tissue is reperfused after, for instance, one hour, a strong acceleration of the tissue damage will occur instead of an improvement. Most likely there are several reasons for this so-called reperfusion paradox, but oxygen radicals formed as a result of the re- appearance of oxygen in previously ischaemic tissue appear to contribute to the damage. The radicals are extremely short-lived and therefore difficult to study directly, and much of the information concerning their formation and effects is inferred from the protective action of various oxygen radical scavengers. Their formation have however also been demonstrated more directly by means of EPR on heart samples. Tissue protection by means of CuZn SOD has been demonstrated in ischaemia- or anoxia-reperfusion models in the kidney.

It is contemplated that EC-SOD and EC-SOD variants might be used in connection with thrombolytic agents such as streptokinase, tissue plasminogen activator, urokinase and variants and variants of these factors.

The source of oxygen radicals in this situation is not completely clear, but the effect of allopurinol seems to indicate that it is partly caused by xanthine oxidase which, by ischaemia, is converted from its xanthine dehydrogenase form to the radical-producing oxidase form. A large amount of hypoxanthine which is the substrate for xanthine oxidase is formed due to purine nucleotide degradation induced by ischaemia. Other sources of superoxide radicals may be activated leukocytes attracted to ischaemia-damaged tissue, prostaglandin synthesis,  $O_2^{\cdot -}$  is a byproduct; and autooxidation of various compounds accumulated in reduced form during ischaemia. The finding concerning ischaemia followed by reperfusion has potentially important clinical applications.

- It may be possible to obtain an excellent effect by reperfusion of tissue in connection with heart infarctions, by the concomitant administration of a SOD such as EC-SOD or a polypeptide of the invention and/or other protective factors against oxygen radicals and thrombolytic factors, e.g. tissue plasminogen activator. The results of the SOD experiments also indicate a possible application in connection with heart surgery and heart transplantations. Analogously, the results of employing an SOD in connection with kidney ischaemia followed by reperfusion indicate that SOD may be employed in connection with kidney transplantations and other organs transplantations such as skin, lung, liver, pancreas, bone tissue transplantations. Ischaemic brain disease is another possible indication.
- 15 In the case of, for instance, burns, immunocomplex formation, and major tissue damage, neutrophilic leukocytes are accumulated in the lungs. Complement activation (C5 a) often seems to mediate the accumulation. The leukocytes seem to be activated and release oxygen radicals, thereby causing lung damage which, for instance, is characterized by increased vessel permeability and lung oedema (adult respiratory distress). In several animal models, SOD and other oxygen radical scavengers have been shown to have a protective effect against lung damage.
- 25 Concerning the central nervous system protective effects of CuZn SOD have been shown against posttraumatic brain oedema.

The endothelium-derived vessel relaxant factor (EDRF) is very sensitive to superoxide, and administration of SOD augments its actions. Superoxide radical production can occur under many circumstances in the body and may cause vasoconstriction and decreased tissue perfusion. Administration of SOD is believed to be able to relieve such vasoconstriction and also to enhance other effects of EDRF such as platelet stabilization.

Acute severe increase in blood pressure leads to functional and morphologic abnormalities in brain arterioles. Prostaglandin synthesis inhibitors and superoxide dismutase is contemplated to protect against the abnormalities. Superoxide release can be detected. Close analysis of the model has led to the conclusion that superoxide radicals are formed as a byproduct during prostaglandin synthesis. The results suggest that tissue damage caused by superoxide radicals released during prostaglandin synthesis may occur in other pathological situations and that SOD may exert a protective action.

In various types of autoimmune disease, such as systemic lupus erythematosus (SLE), systemic sclerosis and rheumatoid arthritis, an increased frequency of chromosomal breaks in lymphocytes has been demonstrated.

The neoplastic transformation of cells is usually divided into two phases, i.e. initiation followed by promotion. In *in vitro* models where initiation has been caused by ionizing radiation, bleomycin, misonidazole and other nitroimidazoles, the oncogenic transformation has been effectively inhibited by the presence of SOD in the medium. It is not necessary for SOD to be present during exposure to the initiating substances which seems to indicate that the enzyme inhibits the subsequent promotion step. Non-toxic doses of xanthine + xanthine oxidase cause promotion in growing cells. Addition of SOD or SOD + catalase inhibits this effect. Phorbol esters are known promoters. In a model in which skin tumors were induced by initiation with a benzanthrane followed by application of a phorbol ester (TPA), local treatment with a lipophilic copper complex with SOD activity strongly reduced tumor formation. The result indicates that, at least in certain cases, superoxide radicals contribute to the promotion of tumor formation and that SOD may protect against this effect.

There is reason to believe that oxygen radicals contribute to the damaging effects of a number of toxic substances such as

bleomycin, adriamycin, alloxan, 6-hydroxydopamine, paraquat, dihydroxyfumaric acid, nitrofurantoin and streptozotocin. In those cases where radical formation takes place in the extracellular space it might be possible to protect by means of injected protective enzyme. Thus, SOD may protect against the diabetogenic activity of alloxan (damages  $\beta$ -cells in the pancreas) *in vitro* and *in vivo*. The damaging effect of alloxan seems therefore to be mediated by the superoxide radical or by other oxygen radicals derived from it. In diabetes mellitus there is an infiltration in the Langerhans' islets by inflammatory cells which potentially may form oxygen radicals. It may therefore be contemplated to protect the  $\beta$ -cells by injections with SODs such as EC-SOD or a polypeptide of the invention at the first onset of diabetes mellitus.

Recombinant EC-SOD C has been found to be more efficient than CuZn SOD in disease models in which the two SODs have been tested in parallel. Thus, recombinant EC-SOD C has been shown to reduce the concentration of oxygen-free radicals in reper-fused rat hearts. The effect of rEC-SOD C in reducing the free radical concentrations was concluded to be at least of the same extent as CuZn SOD. Furthermore, recombinant human EC-SOD C has been shown to reduce myocardial damage in rats subjected to ischemia and 24 hours of reperfusion.

The dosage and timing of polypeptide of the invention injections depend on the half-life of the enzyme in human blood vessels. It may as in rabbits be about 15 hours. The half-life in humans is however probably longer. Assuming first-order kinetics and a half-life of 36 hours, daily injections of 35 mg after an initial injection of 87 mg would therefore result in the same concentration as after the initial injection.

The therapeutic usefulness of the polypeptides of the invention may vary according to the particular disease to be treated. When strong binding to cell-surfaces is advantageous



EC-SOD C or the polypeptides of the invention with even stronger heparin affinity may be most useful, for example in transplantation of organs or if a polypeptide of the invention is to be injected into an inflamed organ.

- 5 In yet other conditions a more limited heparin affinity such as that of a polypeptide having a slightly reduced affinity e.g the polypeptide T216, described herein, may be advantageous. In a situation in which the time of reperfusion cannot be predicted precisely (e.g. thrombolysis of coronary
- 10 artery or artery in other organs) a strong binder such as EC-SOD C injected before reperfusion would bind firmly to endothelium around the body and very little would be available to protect immediately the reperfused organ territory at reperfusion. A polypeptide having a slightly reduced affinity,
- 15 nity, e.g the polypeptide T216 described herein, would in this situation exist in relatively high concentration in the plasma phase, but would still bind endothelium and is also likely to redistribute more rapidly between binding sites in organs. Only EC-SOD variants with a heparin affinity of a
- 20 magnitude which results in elution from a Heparin-Sepharose® column at a NaCl concentration of 0.55 M or higher (under the conditions stated in Example 6) will bind to endothelium *in vivo*. Such variants may be advantageous in conditions where some but variable degree of cell-surface binding is valuable.
- 25 Variants which apparently do not bind heparan sulfate *in vivo*, A and B-types, may finally be useful when a high SOD activity in plasma or the interstitial fluid phase is important.

- For topical treatment, far less of a polypeptide composition
- 30 of the invention as described above would probably be needed. At present, 4-8 mg of CuZn SOD are administered intraarticularly once a week for the treatment of arthritis. EC-SOD which has a far higher molecular weight is likely to remain in the joint for a longer period of time. A similar treatment

protocol or possibly somewhat lower doses will probably be appropriate.

The polypeptide of the invention having the superoxide dismutating property of native EC-SOD may further be used in cosmetics, e.g. to prevent skin ageing and protect against radiation damages. The polypeptide may be in a liposome composition.

#### LEGEND TO FIGURES

Figure 1. C-terminal cDNA sequences and amino acid sequences of native EC-SOD and 12 EC-SOD variants.

Figure 2. Structure of the recombinant WAP/BC-SOD gene and identification of hEC-SOD Transgenic Mice by Southern Analysis.

A. Structure of the WAP/EC-SOD transgene. To direct expression of recombinant human EC-SOD to the mammary gland of transgenic mice a hybrid WAP/EC-SOD gene, pS172, was constructed. This recombinant gene comprises about 2.3 kb of upstream regulatory sequences including the first 24 bp of the transcribed but untranslated WAP sequence in front of the human EC-SOD cDNA. Downstream of the cDNA, a WAP genomic fragment of about 4 kb extending from the SAlI site located in the third exon to the EcoRI site located about 1.6 kb 3' of the last exon was inserted.

B. Southern blot analysis of integrated WAP/EC-SOD transgenes. Tail DNA, 10 µg, from each animal was restricted with samHI, electrophoresed on 1% agarose gel, transferred to membranes and hybridized to a 32P-labelled probe of EC-SOD cDNA. The hybridizing band of 1.8 kb contains the EC-SOD cDNA and 3' WAP sequences. Lane 1 is a control whereas lanes 2-15 each represent an animal.

**Figure 3. Tissue distribution of WAP/EC-SOD mRNA expression in a lactating female of line #68, 11 days after parturition.**

- A) 10  $\mu$ g of total RNA from various tissues was electrophoresed on agarose-formaldehyde gel, transferred to Gene Screen Plus membranes (New England Nuclear) and hybridized with a 32P-labelled EC-SOD cDNA probe. The RNA hybridization results show expression in the lactating mammary gland of a 1 kb major EC-SOD mRNA species; no hybridization to RNA prepared from non-transgenic animals was observed.
- 10 The tissues were mammary gland (Mg), liver (Li), kidney (Ki), spleen (Sp), heart (He), lung (Lu), salivary gland (Sg) and brain (Br). RNA sizes in kb, determined by co-electrophoresis of a RNA ladder (BRL) is indicated to the left.
- B) Northern blots of mammary gland RNA of transgenic line #68 and controls were hybridized with a mouse WAP exon 3 and exon 4 probe, which hybridizes to the same extend to both endogenous WAP mRNA and to recombinant WAP/EC-SOD mRNA. The endogenous WAP mRNA level was found to be significantly higher than the level of recombinant WAP/EC-SOD mRNA. The  $\beta$ -lactoglobulin/EC-SOD transgenic lines were not analysed for mRNA expression.

**Figure 4. Immunoblot of human EC-SOD produced in milk from a transgenic mouse of line #68. Electrophoresis, transfer and development was performed as described in materials and methods. Approximately 400 ng of EC-SOD was applied to each lane. Lanes 1 and 6: Molecular weight markers (116.5, 80, 49.5, 32.5, 27.5 and 18.5 kDa). Lane 2: Recombinant EC-SOD produced in CHO cells. Lanes 3-5: The whey fraction of milk collected at day 4, 10 and 18 post parturition, respectively.**

**Figure 5. The EC-SOD production in milk from a transgenic mouse of line #68 was followed over lactation. Recombinant EC-SOD levels were determined both with ELISA and SOD activity analysis. The scales for these parameters were chosen so**

that EC-SOD with specific activity identical to that of native and CHO produced recombinant human EC-SOD (8.6 ng/U, ref. 13) will form columns for amount of protein and enzymic activity of equal heights, the columns in black show ELISA concentration and the dotted columns show activity. Milk samples were withdrawn daily as indicated.

**Figure 6. Analytical separation of milk derived recombinant EC-SOD on Heparin-Sepharose.** Milk from line #68, containing about 1800 ng EC-SOD was applied to and separated on a Heparin-Sepharose column as described in the Materials and Methods section (-o-o-), (----) the gradient in NaCl.

**Figure 7. Plasma clearance of recombinant human EC-SOD from mouse milk in rabbits.** The figure shows the time-course in plasma of EC-SOD produced in milk of transgenic mice injected intravenously into rabbits, as described in Materials and Methods. The expected plasma concentration was calculated from the amount injected and the plasma volume as determined with  $^{125}\text{I}$ -albumin. The experiments were terminated after 1 ( $\Delta$ ), 5 ( $\blacksquare$ ) or 24 (o) by intravenous injection of a large dose heparin followed by collection of blood sample after 2, 5 and 15 min.

## EXAMPLES

The following examples are intended to illustrate but not to limit the present invention.

Construction of the expression systems of the invention, and  
5 the molecular biological characterization of it, employs standard methods generally known in the art of recombinant DNA. Unless otherwise stated, the methods used are those described by Sambrook et al., 1989 [70] and Ausubel et al., 1991 [71].

## 10 DEFINITIONS

Hybridization of DNA

DNA, e.g. present on nitrocellulose filters, are wetted in 2 x SSC [1 x SSC: 0.15 M NaCl, 0.0015 M Na<sub>3</sub>-citrate, pH 7.0] and placed in a heat-sealed plastic bag with pre-warmed  
15 (67°C) prehybridization solution. Prehybridization takes place for 2 h at 67°C, the bag being gently shaken. The solution is exchanged with pre-warmed (67°C) hybridization solution, a radioactive probe is added and hybridization is carried out at 67°C for 18 h. The bag is gently shaken to  
20 ensure constant movement of the liquid over the nitrocellulose filters. After hybridization, a washing procedure is carried out.

The radioactive probe is prepared by use of known methods, e.g. as described by Sambrook et al. [70], on the basis of  
25 the DNA sequence shown in SEQ ID NO:1 or a part thereof, especially a coding part such as the nucleotides corresponding to amino acids 1-210 or an effective subsequence of the DNA sequence as defined above.

The prehybridization and hybridization solutions used are:  
30 10 x Denhardt's, 4 x SSC, 0.1% SDS, 10 µg/ml polyA, 50 µg/ml of denatured DNA to be analyzed and the denatured (heat)

radioactive probe. The filters are washed in pre-warmed (67°C) solutions: 10 x Denhardt, 2 x SSC, 0.1% SDS for 2 x 15 min. and 1 x SSC, 0.1% SDS for 4 x 15 min. The filters are air-dried and covered with Vita-Wrap, and X-ray film is  
5 exposed to the filters for 3 h to 3 weeks with and without intensifying screens.

#### EXAMPLE 1

##### Construction of transgenes

3 vectors were constructed, pS172, pS315 and pS387.

10 The human EC-SOD cDNA [12] was subcloned as a 1396 bp EcoRI fragment into pUC18. The cDNA was modified in the 5' end by insertion of a HindIII linker 32 bp upstream of the start codon and at the 3' end by insertion of a SalI linker downstream of the translational stop. To facilitate introduction  
15 of human EC-SOD cDNA under control of the WAP promoter, a plasmid harbouring the 7.2 kb EcoRI fragment containing the murine WAP gene [39] was digested with KpnI and a HindIII linker was inserted. The resulting plasmid was digested with HindIII and SalI thereby removing the sequence between KpnI  
20 in exon I and SalI located in exon III of the WAP gene. This fragment was then ligated with the HindIII/SalI human EC-SOD cDNA fragment. The resulting vector, pS172, was then digested with EcoRI and a fragment of about 5.9 kb (Fig. 2) was isolated and purified by electro-elution before injection into  
25 mouse embryos.

A variant of pS172 was also generated. This vector, pS315, has a longer 5'WAP of about 4.5 kb and the translational start of EC-SOD was inserted directly downstream of the KpnI site in exon I of the murine WAP gene. This KpnI site is  
30 located just upstream of the translational start of WAP. The WAP/EC-SOD recombinant gene was isolated, purified by electroelution and injected as described below.

A third variant of expression vector containing 1.8 kb ovine  $\beta$ -lactoglobulin upstream regulatory sequence [40] in front of human EC-SOD cDNA was constructed. The ovine  $\beta$ -lactoglobulin promoter fragment was cloned as a BamHI/PvuII fragment into  
5 BamHI/SmaI digested pUC19. From this plasmid, the promoter fragment was then isolated as a 1.8 kb HindIII/KpnI (partial digestion) fragment. This  $\beta$ -lactoglobulin promoter fragment was then ligated to a human EC-SOD cDNA which was modified by insertion of a KpnI site in front of the start codon. Down-  
10 stream of the EC-SOD stop codon, a murine 4.6 kb WAP genomic fragment was inserted, providing the mRNA processing signals. This 4.6 kb WAP fragment extends from the KpnI site, which was changed to a SalI, in the first exon to the EcoRI site located about 1.6 kb downstream of the last exon. The  $\beta$ -lac-  
15 toglobulin/ECSOD/WAP fragment was isolated from the resulting expression vector, pS387, and injected.

The milk protein gene sequences were kindly provided by Dr. Lothar Hennighausen.

### Injection

20 An EcoRI fragment (Fig. 2) was isolated from the plasmid pS172 and used for injection. Alternatively, the WAP/ECSOD fragment of pS315 or  $\beta$ -lactoglobulin/EC-SOD/WAP fragment of pS387 was isolated and injected. The isolated fragments were injected into the pronucleus of C57Bl/6JxCBA-f2 or  
25 C57Bl/6JxDBA/2J -f2 embryos obtained after priming of donor female mice with 5 IU pregnant mare's serum gonadotropin followed by 5 IU human chorion gonadotropin 48 h later for superovulation of the mice. The C57Bl/6JxCBA-f1 animals were obtained from Bomholtgaard Breeding and Research Centre Ltd.  
30 (Ry, Denmark). The injections were performed using Narishigi hydraulic micromanipulators and a Nikon inverted microscope [41]. After injection the embryos were implanted in pseudo-pregnant C57Bl/6JxCBA-f1 recipients.

## Results

To direct expression of recombinant human EC-SOD to the mammary gland of transgenic mice a hybrid WAP/EC-SOD gene, pS172, was constructed. This recombinant gene comprises about 2.3 kb of upstream regulatory sequences including the first 24 bp of the transcribed but untranslated WAP sequence in front of the human EC-SOD cDNA. Downstream of the cDNA, a WAP genomic fragment of about 4 kb extending from the *SalI* site located in the third exon to the *EcoRI* site located about 1.6 kb 3' of the last exon was inserted (Fig. 2A). To generate WAP/EC-SOD transgenic animals 178 injected ova at one-cell stage were implanted into 7 foster mothers resulting in 19 newborn mice. 14 animals were analyzed and five mice were identified as carrying WAP/EC-SOD sequences (founder animals) using PCR analysis, and the presence and the integrity of the injected DNA was confirmed with Southern blot analysis (Fig. 2B). Three of the founder animals were females and two were males. One female founder was lost. Founder animals were bred and lactating females were generated from all the lines.

The  $\beta$ -lactoglobulin/EC-SOD construct was injected in 160 ova which were implanted into 16 foster mothers. This resulted in birth of 28 mice which were analyzed. Among these, three mice were identified to be transgenic and were analyzed for expression of recombinant protein.

## 25 Conclusion

Five lines of transgenic mice with integrated WAP/EC-SOD gene were identified. Active recombinant EC-SOD was obtained in milk from such animals and expression of the recombinant gene was found predominantly in the mammary gland. The  $\beta$ -lactoglobulin/EC-SOD hybrid gene was integrated into three lines of transgenic mice. This construct was found to mediate very low levels of human ECSOD expression in the mammary gland. The level of active human EC-SOD in milk from  $\beta$ -lactoglobulin/-EC-SOD transgenic mice was below 10 ng/ml whereas the WAP/-



EC-SOD hybrid gene mediated levels up to 0.7 mg/ml. No correlation between expression levels and copy number of the recombinant EC-SOD genes could be observed. This is in agreement with other studies [48, 49].

## 5 EXAMPLE 2

### Early identification of transgenes

Mice having integrated the transgene were identified with PCR analysis of DNA from tail biopsy specimens obtained three weeks after birth of the animals. Biopsies were incubated in  
10 0.6 ml urea lysis buffer containing; 1 M Urea, 50 mM Tris-HCl pH 8.0, 0.1 M NaCl, 0.5% SDS, 5 mM EDTA and 200 mg/ml proteinase K. Samples were incubated overnight at 37°C. PCR analysis was performed in 50 mM KCl, 10 mM Tris-HCl pH 8.4, 1.5 mM MgCl<sub>2</sub>, 0.25 mM dNTP, 2.5 U Taq polymerase (BRL, Gaithersburg, MD), using primers complementary to WAP (5'-CTGTGTGGCCAAGAAGGAAGTTTGT-3') (SEQ ID NO:3),  $\beta$ -lactoglobulin (5'-GCCTGAGGATGAGCCAAGTG-3') (SEQ ID NO:4) and EC-SOD (5'-GTCCAGCGTGGCCGACGGCTGCACCTG-3') (SEQ ID NO:5) sequences,  
15 respectively. Positive results were confirmed by Southern blot analysis using the human EC-SOD cDNA as a probe [42].  
20

## EXAMPLE 3

### Expression of transgenes

Expression of the transgenes were assessed by analyzing RNA and milk from lactating females that were either f<sub>0</sub> animals  
25 or the transgenic f<sub>1</sub> offspring of f<sub>0</sub> males.

### RNA isolation and analysis

Total RNA from various tissues of transgenic lactating female mice was isolated [42]. Samples of 10  $\mu$ g total RNA was fractionated on MOPS/formaldehyde agarose gels and transferred to  
30 Gene Screen plus membranes (New England Nuclear, Boston, MA),

and subsequently hybridized to a  $^{32}\text{P}$ -labelled probe according to the supplier's protocol (Amersham, U.K.).

The RNA hybridization results show expression in the lactating mammary gland of a 1 kb major EC-SOD mRNA species (Fig. 3A). This is in agreement with the expected mRNA size. No hybridization was observed to RNA prepared from non-transgenic animals.

The tissue distribution of recombinant WAP/EC-SOD expression was investigated in duplicate mice, of the transgenic lines. Analysis of RNA prepared from various tissues (Fig. 3A) showed that abundant expression of recombinant EC-SOD was restricted to the lactating mammary gland. However, low levels of recombinant EC-SOD expression was also detected in brain. In additional experiments, where the same WAP regulatory sequences were used to direct expression of other human cDNAs, no recombinant gene expression was detected in the brain.

Northern blots of mammary gland RNA of transgenic line #68 and controls were hybridized with a mouse WAP exon 4 probe, which hybridizes to both endogenous WAP mRNA and to recombinant WAP/EC-SOD mRNA. The endogenous WAP mRNA level was found to be significantly higher than the level of recombinant WAP/EC-SOD mRNA (data not shown). The  $\beta$ -lactoglobulin/EC-SOD transgenic lines were not analyzed for mRNA expression.

#### EXAMPLE 4

##### Milk collection.

Mice were anaesthetized with 0.017 ml Avertin (2.5%) per g body weight [41] and given 0.2 IE oxytocin (Partocon, Ferring, Lund, Sweden) intraperitoneally ten minutes before milk collection. The milk was collected using a milk apparatus which collected the milk by continuous suction into chilled

1.5 ml Eppendorf tubes. After collection the milk was immediately frozen in  $-70^{\circ}\text{C}$ .

#### EXAMPLE 5

##### Determination of EC-SOD in mouse milk

##### 5 Partial purification of EC-SOD produced in transgenic mice

Milk collected from mice were diluted ten times with distilled water and thereafter centrifuged at  $8000 \times g$  for 5 min. The lipid layer at the surface was carefully removed and the supernatant was collected. The skimmed milk was diluted 1:1  
10 with 50 mM sodium phosphate, 0.5 M NaCl, pH 7.0 (buffer A) and thereafter applied to a column with a monoclonal antibody directed against native human EC-SOD coupled thereto. The column was washed with buffer A and eluted with 50 mM AMP-HCl, 1 M KSCN, pH 9.0. The eluate was ultrafiltrated and  
15 transferred to distilled water and stored frozen at  $-20^{\circ}\text{C}$ . The protein concentration of collected fractions was measured using the method of Bradford [43].

##### Enzyme-linked immunosorbent assay (ELISA) for human EC-SOD

Quantitation of human EC-SOD was made using the double antibody sandwich ELISA method. Microtiter plates (Nunc; Nunc, Roskilde, Denmark) were coated with  $100 \mu\text{l}$  per well of a solution containing  $16 \mu\text{g/ml}$  of polyclonal rabbit anti-EC-SOD IgG antibodies in 50 mM  $\text{Na}_2\text{CO}_3$ , pH 9.6. After 2 h incubation at room temperature, the wells were washed with blocking  
25 buffer (10 mM Na phosphate, pH 7.4, 140 mM NaCl, 0.1 % wt/vol Tween 20, and 0.5 % BSA) and then blocked overnight with  $300 \mu\text{l}$  blocking buffer. For analysis,  $50\text{-}\mu\text{l}$  samples, diluted if necessary with blocking buffer, were added to each well and incubated for 2 h. The wells were then washed with blocking  
30 buffer, and  $50 \mu\text{l}$   $3 \mu\text{g/ml}$  monoclonal anti-EC-SOD antibody B6,H6 which is described in, and produced according to, Example 15 of WO 87/01387 and  $50 \mu\text{l}$  peroxidase-conjugated

rabbit anti-mouse IgG (DAKOPATTS, Copenhagen, Denmark), both dissolved in blocking buffer, added in that order. After a further 2 h, the wells were washed with blocking buffer and then developed for 10 min with 100  $\mu$ l 3.7 mM O-phenylene-diamine and 0.4 mM H<sub>2</sub>O<sub>2</sub> in 100 mM Na citrate, pH 5.0. After addition of 100  $\mu$ l 0.5 M H<sub>2</sub>SO<sub>4</sub>, the absorbance at 492 nm was determined in an ELISA processor II (Hoechst Behring, Marburg, FRG). The assay was standardized with human umbilical cord EC-SOD C. EC-SOD concentrations down to ~0.25 ng/ml could be determined.

The production of EC-SOD determined with ELISA varied a lot between transgenic lines but overall highest in the WAP transgenes (Table 1). One line, #68, displayed a very high productivity, around 0.7 mg/ml.

Table 1.

**Production of human EC-SOD in milk of transgenic mice**

	Vector	Transgenic line	EC-SOD (ng/ml)
20	pS172	65	90
		66, 67	UD <sup>a</sup>
		68	710000
	pS315	90	15
	pS387	607, 702	UD
		716	8
25	None	Control	0-0.75

aUD, Undetectable, within the range of controls.

SDS-PAGE and Immunoblotting for the presence of human EC-SOD

SDS-gels were run in a Midget Electrophoresis Unit (Pharmacia-LKB) according to the discontinuous buffer system described by Laemmli [46]. Gels were blotted in 48 mM Tris, 39 mM glycine, 1.3 mM SDS, 20 % methanol, in a Trans Blot unit

(Bio-Rad, Richmond, CA) under constant voltage (0.8 mA/cm<sup>2</sup>). EC-SOD was detected by the use of polyclonal antibodies prepared in rabbits using the native protein for immunization and immobilized EC-SOD for affinity purification. Alkaline phosphatase anti rabbit IgG (Dakopatts, Copenhagen, Denmark) was used as secondary antibody.

Milk was analyzed by SDS/PAGE and immunoblotting for the presence of human EC-SOD (Fig. 4). With this method, human EC-SOD was detected in milk from the WAP/EC-SOD line #68, but was not detected in milk from the other lines or in milk from non-transgenic animals.

#### EXAMPLE 6

##### Characterization of EC-SOD produced in transgenic mice

Native EC-SOD and recombinant EC-SOD expressed in CHO-cells was obtained from Symbicom AB (Umeå, Sweden).

##### Quantification of EC-SOD activity

The superoxide dismutase activity of samples was determined by a direct spectrophotometric method employing KO<sub>2</sub> [44], as slightly modified in [45]. In brief, the basis of the method is that SOD is determined in terms of its ability to catalyze the disproportionation (decay) of O<sub>2</sub><sup>•-</sup> in alkaline aqueous solution. At alkaline pH and low O<sub>2</sub><sup>•-</sup> concentration, the O<sub>2</sub><sup>•-</sup> radical is stable enough to be studied in common UV-vis spectrophotometers, using the broad absorbance maximum at 245-250 nm. Thus the assay was conducted at pH 9.5 and the disproportionation studied directly in a spectrophotometer. One unit in the assay is defined as the activity that brings about a disproportionation of O<sub>2</sub><sup>•-</sup> at a rate of 0.1 sec<sup>-1</sup> in 3 ml of buffer. Under these conditions, one unit of enzyme corresponds to 8.6 ng native or recombinant human EC-SOD C [13].

This assay is 40 times more sensitive (i.e. the units correspond to 40 times less enzyme) than the original xanthine oxidase - cytochrome C assay [9].

### Results

5 The SOD activity of control milk was about a hundred U/ml, why enhancements in activity in the milk samples with low EC-SOD production not could be distinguished with certainty. However, in line #68 the SOD activity in the milk was very high (Fig. 5), and the specific activity (U/ng EC-SOD) was  
10 apparently identical to that of native and CHO-produced recombinant EC-SOD [13]. Production of recombinant EC-SOD was stable throughout lactation.

### Determination of apparent molecular masses by means of size exclusion chromatography

15

Apparent molecular weight of EC-SOD was determined by gel chromatography, at 4°C using a LKB HPLC apparatus (Pharmacia LKB Biotechnology Inc.). The samples containing human EC-SOD variants were applied to a Sepharyl S-300 column (Pharmacia  
20 LKB Biotechnology Inc.) (1.6 x 89 cm), equilibrated and eluted with 10 mM potassium phosphate, pH 7.4, containing 0.15 M NaCl, at a flow rate of 19.8 ml per hour. The effluent was collected in 1.35 ml fractions and the EC-SOD content determined by ELISA as described above. The column was cali-  
25 brated with IgG 156 (kDa) (Sigma Chemical Co., St. Louis, USA), bovine serum albumin (67 kDa) (Sigma Chemical Co., St. Louis, USA) and carbonic anhydrase (29 kDa) (Sigma Chemical Co., St. Louis, USA). The calibration curve was constructed by means of plotting the log molecular weights of the cali-  
30 brators versus their elution volumes.

The EC-SOD produced in milk from transgenic mice eluted from Sephacryl S-300 size exclusion chromatography column at a position corresponding to the molecular mass 155 kDa (data not shown). The apparent molecular mass of the recombinant

EC-SOD subunit from mouse milk was similar to CHO produced recombinant EC-SOD. Some minor discrepancies were observed. Native EC-SOD of human plasma, and recombinant CHO-produced EC-SOD, chromatographed in parallel, eluted at virtually identical elution volumes. Recombinant EC-SOD from transgenic mice showed approximately 5% protein of higher mers than tetramers. This value is close to that of native human EC-SOD but considerably lower than that of recombinant EC-SOD from CHO cells [47]. The relative amount of dimers was higher in milk derived EC-SOD (Fig. 4). The proportion of the multimers also increased in samples from the later stages of lactation.

#### Glycosylation pattern.

Recombinant EC-SOD produced by CHO cells and mouse mammary glands and native human EC-SOD were stained by PAS and found to contain oxidizable carbohydrate moieties. A characterization of the glycans was made by analyzing the lectin binding properties (Table 2).

The binding of the different SODs to lectins was analyzed using the lectin-link kit (Gibzyme, Cambridge, MA). This contained the following biotinylated lectins: Concanavalin A (conA), wheat germ agglutinin (WGA), *Ricinus communis* agglutinin (RCA), *Datura stramonium* agglutinin (DSA) and *Sambucus nigra* agglutinin (SNA). The proteins were first run on SDS-PAGE and thereafter electroblotted and developed using the different lectins.

Both CHO and mouse milk derived recombinant EC-SOD bound RCA weaker than the native protein. This indicates that the native protein may contain terminal galactose which are absent or blocked by sialic acid in the recombinant proteins. The three EC-SODs differ with respect to SNA binding. Only milk derived recombinant EC-SOD was found to bind this lectin, indicating that EC-SOD produced in transgenic mice has terminal sialic acid residue(s) attached to galactose in  $\alpha 2,6$  (or  $\alpha 2,3$ ) position.

Table 2

## Lectin binding of native and recombinant human EC-SOD

Comparison of the lectin binding properties of native human umbilical cord EC-SOD (n EC-SOD), recombinant EC-SOD produced in CHO cells (CHO EC-SOD) and EC-SOD produced in milk from transgenic mice (TG EC-SOD). The binding is divided into four different strengths: no (-), weak (+), medium (++), and strong (+++). The lectins are abbreviated as described in materials and methods.

	RCA	ConA	SNA	DSA	WGA
n EC-SOD	+++	++	-	+	-
CHO EC-SOD	+	+++	-	++	+
TG EC-SOD	+	++	++	+	++

Plasma clearance of EC-SOD in rabbits

Recombinant EC-SOD from mouse milk was partially purified by chromatography on Heparin-Sepharose equilibrated with 50 mM Na phosphate, pH 7.4. Sound EC-SOD was eluted with a gradient of NaCl in the same buffer. The central 2/3 of the high heparin-affinity fraction (cf. Fig. 6) was collected and used for plasma clearance studies. About 10 µg of EC-SOD per kg body weight, dissolved in 50 mM potassium phosphate, pH 7.4, containing 0.2% bovine serum albumin, was injected into ear veins of chinchilla ram rabbits, weights 3-4 kg. Blood samples were tapped at times indicated in Fig. 7 into tubes containing EDTA as an anti-coagulant. The experiments were terminated at 1, 6 or 24h, cf. Fig. 7, by intravenous injection of heparin, 2500 IU/kg body weight, followed by tapping of blood samples at 2, 5 and 15 min thereafter. The plasma volumes varied between 34 and 38 ml/kg body weight. The rabbits were used only once.



After intravenous injection milk derived recombinant EC-SOD was rapidly, to about 97%, sequestered from the blood plasma (Fig. 7). This sequestering is apparently due to binding to heparan sulphate proteoglycan in the glycocalyx of endothelial cell surfaces [3, 4, 15, 17, 18]. The decline thereafter was slow. Injection of large doses of heparin at 1, 5 and 24 hours lead to prompt release of the enzymes to plasma. This behaviour is very similar to that previously found with CHO produced recombinant EC-SOD.

10 However, CHO produced EC-SOD displayed a somewhat slower elimination from the circulation.

#### Analytical separation of EC-SOD on Heparin-Sepharose

EC-SOD was separated by Heparin-Sepharose chromatography using a Pharmacia FPLC apparatus, at room temperature. The columns contained 1 ml Heparin-Sepharose (Pharmacia Laboratory Separation Division, Uppsala, Sweden) with 15 mM Na cacodylate/50 mM NaCl, pH 6.50, as equilibration buffer and eluent. The absorbance at 280 nm was monitored on the eluent. The samples were applied at 5 ml per hour and when the absorbance at 280 nm approached the baseline, bound components were eluted with a linear gradient of NaCl in the buffer (0-1 mol/l) at 9 ml/h. The effluent was collected in 0.65 ml fractions and EC-SOD determined by enzyme-linked immunosorbent assay (ELISA), which method is described below. The chloride content of EC-SOD was determined using a standard chloride titrator (American Instrument Co, Inc, Md, USA).

Before application the solvents of the culture media were exchanged to 15 mM sodium cacodylate/50 mM NaCl, pH 6.50, on a Filtron Omega cell membrane concentration system. In general 1-2 ml of culture medium was applied to the Heparin-Sepharose® column.

Upon chromatography on Heparin Sepharose milk derived recombinant EC-SOD produced in transgenic mice displays a major

peak eluting at about 0.54 m NaCl, a concentration identical to that seen with native and CHO produced recombinant EC-SOD [19], (Fig. 7). There was, however, some material with reduced and aberrant heparin affinity.

- 5 The recombinant EC-SOD produced in milk was apparently structurally identical to the previously studied native plasma and CHO-produced recombinant human EC-SODs. The apparent molecular masses were indistinguishable upon gel exclusion chromatography as were the subunits upon SDS/PAGE-immunoblotting.
- 10 The specific enzymic activity was virtually identical to those of native and CHO-produced recombinant EC-SOD. The major part of the milk produced EC-SOD displayed a heparin affinity identical to that of the previously studied EC-SODs. The heparin-binding domain of EC-SOD is highly susceptible to
- 15 proteolytic truncation [20]. The present investigation indicated that only minor such cleavage occurred despite the presence of proteolytic activity in mouse milk [50].

- As judged from the lectin-affinity investigation, there were differences in the oligosaccharide moieties between all the
- 20 studied human EC-SOD types, native enzyme from umbilical cords, and recombinant produced by CHO cells and mouse mammary glands. However, the finding does not necessarily indicate that the recombinant EC-SOD's contain oligosaccharide moieties that do not exist bound to EC-SOD in the human body.
- 25 The oligosaccharides are known to be heterogeneous, even as produced by a single cell type [51]. The umbilical cord native EC-SOD studied is probably mainly produced by fibroblast. However, EC-SOD is also secreted by glia cells [52], smooth muscle cells (unpublished) and possibly other not yet
- 30 identified cell types. It is well known that the glycosylation of a protein may vary widely when it is produced by different cell types in the human body [53].

- Finally, the recombinant EC-SOD from mouse milk was found to behave similarly to native and recombinant CHO-produced
- 35 EC-SOD [15, 18] after intravenous injection into rabbits.

There was a prompt binding to the vascular endothelium as well as a rapid release after injection of heparin. Possibly, the mouse milk produced EC-SOD was a little more rapidly eliminated from the circulation than the previously studied  
5 forms.

#### EXAMPLE 7

##### Cloning, sequencing and organization of genomic human EC-SOD DNA

The human EC-SOD gene is identified and isolated from a human  
10 genomic library.

Recombinant phages containing EC-SOD genomic sequences are identified by hybridization to <sup>32</sup>P-labelled EC-SOD cDNA fragments. These labelled EC-SOD sequences that are used as probes are the entire EC-SOD cDNA, and in addition, isolated  
15 fragments from both the 5'- and 3'- end of the EC-SOD cDNA. The use of these probes in various combinations allows the identification of the recombinant EC-SOD phages that contain the entire transcribed region. This analysis of the recombinant phages and the EC-SOD gene is even more refined using  
20 various combinations of synthetic oligonucleotides as primers in polymerase chain reaction experiments.

Identified recombinant phages containing the transcribed region of the human EC-SOD gene are plaque purified. DNA is prepared from the purified phages and the EC-SOD genomic  
25 sequences are isolated by restriction enzyme digestion and agarose electrophoresis. The EC-SOD fragments are cloned into pUC plasmids for more detailed restriction mapping, PCR analysis and sequence analysis.

The result shows that EC-SOD has at least one intron preceding the EC-SOD open reading frame. The EC-SOD open reading  
30 frame is continuous in one exon.

To achieve high production of recombinant human EC-SOD in milk from transgenic animals the transcribed intron containing part of the human EC-SOD gene is inserted in a vector, under transcriptional control of mammary epithelial cell specific regulatory sequences derived from a milk protein gene. Transgenic animals harbouring this recombinant gene are generated and identified as described above.

#### EXAMPLE 8

##### In vitro maturation, fertilization and culture of bovine oocytes

Immature oocytes are obtained in large quantity (400-600/day) by aspirating follicles of ovaries obtained at abattoirs. Immature oocytes are cultured for a period *in vitro* before they are competent to be fertilized. Once "matured", oocytes are fertilized with sperm which has also been matured, or "capacitated" *in vitro*. The pronuclei of the fertilized oocyte is then injected with the transgene coding for the expression and secretion of human EC-SOD. Zygotes resulting from this *in vitro* fertilization and microinjection are then cultured to the late morula or blastocyst stage (5-6 days) in medium prepared, or "conditioned" by oviductal tissue. Blastocysts are then transferred non-surgically to recipient bovine species for the balance of gestation or analyzed for integration of the transgene as described herein.

##### 25. *In vitro* maturation (IVM)

Ovaries are obtained immediately after slaughter at local abattoirs and oocytes are recovered. Alternatively, oocytes are obtained from living bovine species by surgical, endoscopic, or transvaginal ultrasonic approaches. In all cases, oocytes are aspirated from ovarian follicles (2-10 mm diameter). After washing, oocytes are placed in a maturation medium such as a medium consisting of M199 supplemented with

10% fetal calf serum, and incubated for 24 hours at 39°C [72].

#### *In vitro* fertilization (IVF)

Matured oocytes are fertilized with either fresh or thawed sperm. Sperm is prepared for fertilization by first obtaining a population of sperm enriched for motility by a "swim-up" separation technique [73]. Motile sperm is then added to a fertilization medium, consisting of a modified Tyrode's solution [73] supplemented with heparin to induce sperm capacitation [74]. Capacitation constitutes the final sperm maturation process which is essential for fertilization. Sperm and oocytes are co-cultured for 18 hours. A useful feature of this IVF method is that (in the case of frozen sperm) consistent, repeatable results are obtained once optimal fertilization conditions for a particular ejaculate have been defined [73].

#### *In vitro* culture (IVC)

Conventional culture systems, which support development of murine, rabbit, or human ova, do not support development of bovine embryos past the 8-16 cell stage. This problem has been overcome by pre-conditioning culture media with oviductal tissue. Oviduct-conditioned medium will support bovine embryos past the 8-16 cell stage to the blastocyst stage *in vitro* [62].

Bovine embryos did not yield to attempts to culture them *in vitro* past the 8-16 cell "block" until Camous et al. [63] demonstrated cleavage to 216 cells when embryos were co-cultured with trophoblastic tissue.

The co-culture procedure was extended to oviductal tissue, based on the ability of homo- or hetero-oviducts to support development from zygote to blastocyst. Thus, bovine embryos co-cultured with oviductal tissue, or in medium conditioned

by oviductal tissue, developed from zygote to blastocyst in vitro [62, 75]. Blastocysts have been produced in this system after superovulation and artificial insemination, or by in vitro maturation (IVM), and fertilization (IVF) of immature  
 5 oocytes. Blastocysts produced in this fashion resulted in pregnancies and live calves after transfer to recipient animals. The results obtained were as follows:

	Step	Efficiency (%)	Number (per 100)
10			
	IVM	90	90
	IVF	80	72
	IVC	30	22
	Embryo transfer	50	11
15	(% pregnant)		

Therefore, from an initial daily harvest of 500 oocytes, it is expected that approximately 55 pregnancies will result.

#### Preparation of oviduct tissue

#### 20 Co-culture and conditioned medium

1. Obtain oviducts after slaughter or by salpingectomy.
2. Harvest luminal tissue by scraping intact oviduct gently with a glass slide.
3. Wash tissue 5 times in 10 ml modified tyrodes-hepes  
 25 solution [74].
4. Resuspend final tissue pellet in M199 + 10% fetal calf serum at a ratio of 1 volume tissue:50 volumes of media.
5. Tissue suspension can be used for embryo co-culture.

6. Alternatively, media may be conditioned for 48 hours; after centrifuging the suspension, the supernatant may be used as embryo culture medium. Conditioned medium may be stored at -70°C, if desired. Conditioned medium should be used at full strength for embryo culture (no dilution) [75].

#### EXAMPLE 9

##### Generation of transgenic rabbits

- Non-superovulated female rabbits are egg-donors. 19.5 hours after mating the eggs are flushed from their oviducts. An average of 8-10 eggs per donor are recovered. The eggs are analyzed for being morphologically intact, fertilized and revealing clearly a visible pronucleus. About 70 to 80% are found to be usable for microinjection.

##### 15 Injection

One or more of the transgene coding for the expression and secretion of human EC-SOD is/are then injected into the pronuclei of the fertilized eggs. Successful pronuclear injection are monitored by the swelling of the pronucleus.

- 20 After injection, the eggs are cultivated for several hours until the majority reach the two cell stage.

Flushing, culturing and injection are carried out in a medium consisting of phosphate-buffered saline (PBS) supplemented with calf serum.

##### 25 Transference of transgenic eggs

The recipient animals are virgin foster mothers which have been synchronised with the donor animals by a single i.m. injection of FSH releasing hormone. Between 8 and 10 eggs are transferred surgically in general anaesthesia to the oviducts

of the pseudopregnant foster mothers. The abdominal cavity may be opened by flank incisions on both sides and the eggs transferred into the oviducts in 10  $\mu$ l of the culture medium.

The virgin foster mothers are between 4 and 12 months.

- 5 The percentage of pregnancies of the recipient animals are about 60 percent which corresponds to the pregnancy rate observed in natural matings.

The transgenic animals are identified by Southern blot hybridisation of DNA extracted from ear biopsies of the newborn  
10 rabbits. The overall frequency of transgenic rabbits among newborns is about 9.5%.

All of the fertile transgenic animals are mated at an age of 3.5 months and are found to transmit the transgene to their progeny in a Mendelian fashion.



## REFERENCES

1. Marklund, S.L. (1982) *Proc. Natl. Acad. Sci. USA* 79, 7634-7638.
2. Marklund, S.L., Holme, E. and Hellner, L. (1982) *Clin. Chim. Acta* 126, 41-51.
3. Karlsson, K. and Marklund, S.L. (1987) *Biochem. J.* 242, 55-59.
4. Karlsson, K. and Marklund, S.L. (1988) *Biochem. J.* 255, 223-228.
5. Marklund, S.L., Bjelle, A. and Elmqvist, L.-G. (1986) *Ann. Rheum. Dis.* 45, 847-851.
6. Marklund, S.L. (1984) *J. Clin. Invest.* 74, 1398-1403.
7. Marklund, S.L. (1984) *Biochem. J.* 222, 649-655.
8. Sandstrom, J., Karlsson, K., Edlund, T. and Marklund, S.L. (1993) *Biochem. J. In.*, 294, 853-857
9. McCord, J.M. and Fridovich, I. (1969) *J. Biol. Chem.* 244, 6049-6055.
10. Keele, B.B., McCord, J.M. and Fridovich, I. (1970) *J. Biol. Chem.* 245, 6170-6181
11. Weisiger, R.A. and Fridovich, I. (1973) *J. Biol. Chem.* 248, 4793-4796.
12. Hjalmarsson, K., Marklund, S.L., Engstrom, A. and Edlund, T. (1987) *Proc. Natl. Acad. Sci. USA* 84, 6340-6344.
13. Tibell, L., Hjalmarsson, K., Edlund, T., Skogman, G., Engstrom, A. and Marklund, S.L. (1987) *Proc. Natl. Acad. Sci. USA* 84, 6634-6638.
14. Stromqvist, M., Holgersson, J. and Samuelsson, S. (1991) *J. Chromatogr.* 548, 293-301.
15. Karlsson, K. and Marklund, S.L. (1988) *J. Clin Invest.* 82, 762-766.
16. Karlsson, K., Lindahl, U. and Marklund, S.L. (1988) *Biochem. J.* 256, 29-33.
17. Marklund, S.L. and Karlsson, K. (1989) *Lab. Invest.* 60, 659-666.
18. Karlsson, K., Edlund T., Sandstrom, J., Edlund, A. and Marklund, S.L. (1993) *Free Radic. Biol. Med.* 14, 185-190.

19. Sandstrom, J.M., Carlsson, L., Marklund, S.L. and Edlund, T. (1992) *J. Biol. Chem.* 267, 18205-18209.
20. Karlsson, K., Edlund, A., Sandstrom, J. and Marklund, S.L. (1993) *Biochem. J.* 290, 623-626.
- 5 21. Ferrari, R., Ceconi, C., Curello, S., Ghielmi, S. and Albertini, A. (1989) *Pharmacol. Res.* 21, 57-65.
22. Schoenberg, M.H. and Beger, H.G. (1990) *Chem. Biol. Interact.* 76, 141-161.
23. Sanfey, H., Bulkley, G.B. and Cameron, J.L. (1984) *Ann. Surg.* 200, 405-412.
- 10 24. Bannister, J.V., Bannister, W.H. and Rotilio, G. (1987) *CRC Crit. Rev. Biochem.* 22, 111-179.
25. Omar, B.A., Flores, S.C. and McCord, J.M. (1992) *Advances Pharmacol.* 23, 109-161.
- 15 26. Johansson, M., Deinum, J., Marklund, S.L. and Sjoquist, P.-O. (1990) *Cardiovasc. Res.* 24, 500-503.
27. Erlansson, M., Bergqvist, D., Marklund, S.L., Persson, N.H. and Svensjo, E. (1990) *Free Radic. Biol. Med.* 9, 59-65.
- 20 28. Sjoquist, P.-O., Carlsson, L., Jonasson, G., Marklund, S.L. and Abrahamsson, T. (1991) *J. Cardiovasc. Pharm.* 17, 678-683.
29. Sjoquist, P.-O. and Marklund, S.L. (1992) *Cardiovasc. Res.* 26, 347-350.
- 25 30. Abrahamsson, T., Brandt, U., Marklund / Sjoquist, P.-O. (1992) *Circ. Res.* 70, 264-271.
31. Hatori, N., Sjoquist, P.-O., Marklund, S.L., Pehrsson, S.K. and Ryden, L. (1992) *Free Radic. Biol. Med.* 13, 137-142.
- 30 32. Hatori, N., Sjoquist, P.-O., Marklund, S.L. and Ryden, L. (1992) *Free Radic. Biol. Med.* 13, 221-230.
33. Wahlund, G., Marklund, S.L. and Sjoquist, P.-O. (1992) *Free Rad. Res. Comm.* 17, 41-47.
34. Hennighausen, L. (1990) *Prot. Expr. Purif.* 1, 3-8.
- 35 35. Hennighausen, L., Ruiz, L. and Wall, R. (1990) *Curr. Opinion Biotech.* 1, 74-78.

36. Wilmut, A., Archibald, A.L., McClenaghan, M., Simons, J.P., Whitelaw, C.B.A. and Clark, A.J. (1991) *Experientia* 47, 905-912.
37. Simons, J.P., McClenaghan, M. and Clark, A.J. (1987)  
5 *Nature* 328, 530-532.
38. Wall, R.J., Pursel, V.G., Shamay, A., McKnight, R.A., Pittius, C.W. and Hennighausen, L. (1991) *Proc. Natl. Acad. Sci. USA* 88, 1701-1705.
39. Campbell, S.M., Rosen, J.M., Hennighausen, L.G.,  
10 Strech-Jurk, U. and Sippel, A.E. (1984) *Nucl. Acid Res.* 12, 8685-8697.
40. Harris, S., Ali, S., Anderson, S., Archibald, A.L. and Clark, A.J. (1988) *Nucl. Acid Res.* 16, 10379-10380.
41. Hogan, B., Constantini, F. and Lacy, E. (1986) in  
15 *Manipulating the mouse embryo. A Laboratory Manual.* Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y.
42. Current Protocols in Molecular Biology (1992) (Ausubel, F.M., Brent, R., Kingston, R.E., Moore, D.D., Seidman, J.G., Smith, J.A., Struhl, K. eds.) John Wiley & Sons, New York, N.Y.
- 20 43. Bradford, M.M. (1976) *Anal. Biochem.* 72, 248-254.
44. Marklund, S.L. (1976) *J. Biol. Chem.* 251, 7504-7507.
45. Marklund, S.L. (1985) in *Handbook of Methods for Oxygen Radical Research* (Greenwald, R.A. ed.) pp. 249-255, CRC  
25 Press Boca Raton, FL.
46. Laemmli, U.K. (1970) *Nature* 227, 680-685.
47. Stromqvist, M. (1993) *J. Chromatogr.* 621, 139-148
48. Palmiter, R.D. and Brinster, R.L. (1986) *Annu. Rev. Genet.* 20, 465-499.  
30
49. Pittius, W.C., Hennighausen, L., Lee, E., Westphal, H., Nicols, E., Vitale, J. and Gordon, K. (1988) *Proc. Natl. Acad. Sci. USA* 85, 5874-5878.
50. Wilkins, T.D. and Velander, W. (1992) *J. Cell. Biochem.*  
35 49, 333-338.
51. Rademacher, R.W., Parekh, R.B. and Dwek, R.A. (1988) *Ann. Rev. Biochem.* 57, 785-838.
52. Marklund, S.L. (1990) *Biochem. J.* 266, 213-219.

53. Moss, D.W. and Whitaker, K.B. (1985) *Enzyme* 34, 212-216.
54. Oury, T.D., Ho, Y.-S., Piantadosi, C.A. and Crapo, J.D. *Proc. Natl. Acad. Sci. USA*, 89, 9715-9719
- 5 55. Rubnitz, J. and Subramani, S. (1984) *Mol. Cell. Biol.* 4, 2253-2258
56. Brinster, R.L., Allen, J.M., Behringer, R.R., Gelinas, R.E. and Palmiter, R.D. (1988) Introns increase transcriptional efficiency in transgenic mice. *Proc. Natl. Acad. Sci. USA* 85, 836-840
- 10 57. Jaenisch, R. (1976) *Proc. Natl. Acad. Sci. USA* 73, 1260-1264
58. Gordon et al. (1984) *Methods in Enzymology*, 101, 414
59. Hammer et al. (1985) *Nature*, 315, 680
- 15 60. Gandolfi et al. (1987) *J. Reprod. Fert.* 81, 23-28
61. Rexroad et al. (1988) *J. Anim. Sci.* 66, 947-953
62. Eyestone, W.H. et al. (1989) *J. Reprod. Fert.* 85, 715-720
63. Camous et al. (1984) *J. Reprod. Fert.* 72, 779-785
- 20 64. Heyman, Y. et al. (1987) *Theriogenology* 27, 5968
65. Williams et al. (1984) *Theriogenology* 22, 521-531
66. Prather et al. (1987) *Biol. Reprod.* 37, 859-866
67. Roble et al. (1987) *J. Anim. Sci.* 64, 642-664
68. Bongso et al. (1975) *Vet. Res.* 96, 124-126;
- 25 69. Rumsey et al. (1974) *J. Anim. Sci.* 39, 386-391
70. Sambrook, J., Fritsch, E.F. and Maniatis, T.E. *Molecular Cloning, a laboratory manual*. 2nd ed., Cold Spring Harbor Laboratory Press, 1989
71. Ausubel, F.M., Brent, R., Kingston, R.E., Moore, D.D., Seidman, J.G., Smith, J.A., Struhl, K. (eds.), *Current Protocols in Molecular Biology*, John Wiley & Sons, New York, 1991
- 30 72. Sirard et al. (1988) *Biol. Reprod.* 39, 546-552).
73. Parrish et al. (1986) *Theriogenology* 25, 591-600
- 35 74. Parrish et al. (1988) *Biol. Reprod.* 38, 1171-1180
75. Eyestone, W.H. (1989) "Factors affecting the development of early bovine embryos *in vivo* and *in vitro*." Ph.D. Thesis, University of Wisconsin

## SEQUENCE LISTING

(1) GENERAL INFORMATION:

(i) APPLICANT:

(A) NAME: SYMBICOM AB  
(B) STREET: PO BOX 141  
(C) CITY: UMEA  
(E) COUNTRY: SWEDEN  
(F) POSTAL CODE (ZIP): S-901 24

## (ii) TITLE OF INVENTION: Production of EC-SOD

(iii) NUMBER OF SEQUENCES: 31

(iv) **COMPUTER READABLE FORM:**

(A) MEDIUM TYPE: Floppy disk  
(B) COMPUTER: IBM PC compatible  
(C) OPERATING SYSTEM: PC-DOS/MS-DOS  
(D) SOFTWARE: PatentIn Release #1.0, Version #1.25 (EPO)

(2) INFORMATION FOR SEQ ID NO: 1:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 669 base pairs  
(B) TYPE: nucleic acid  
(C) STRANDEDNESS: single  
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(vi) ORIGINAL SOURCE:

(A) ORGANISM: Homo sapiens

(ix) **FEATURE:**

(A) NAME/KEY: CDS  
(B) LOCATION: 1..669

(ix) **FEATURE:**

(A) NAME/KEY: mat\_peptide  
(B) LOCATION: 1..666  
(D) OTHER INFORMATION: /product= "native EC-SOD"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 1:

TGG ACG GGC GAG GAC TCG GCG GAG CCC AAC TCT GAC TCG GCG GAG TGG 48  
Trp Thr Gly Glu Asp Ser Ala Glu Pro Asn Ser Asp Ser Ala Glu Trp  
1 5 10 15

ATC CGA GAC ATG TAC GCC AAG GTC ACG GAG ATC TGG CAG GAG GTC ATG 96  
Ile Arg Asp Met Tyr Ala Lys Val Thr Glu Ile Trp Gln Glu Val Met  
20 25 30

70

CAG CGG CGG GAC GAC GAC GGC ACG CTC CAC GCC GCC TGC CAG GTG CAG Gln Arg Arg Asp Asp Asp Gly Thr Leu His Ala Ala Cys Gln Val Gln 35 40 45	144
CCG TCG GCC ACG CTG GAC GCC GCG CAG CCC CGG GTG ACC GGC GTC GTC Pro Ser Ala Thr Leu Asp Ala Ala Gln Pro Arg Val Thr Gly Val Val 50 55 60	192
CTC TTC CGG CAG CTT GCG CCC CGC GCC AAG CTC GAC GCC TTC TTC GCC Leu Phe Arg Gln Leu Ala Pro Arg Ala Lys Leu Asp Ala Phe Phe Ala 65 70 75 80	240
CTG GAG GGC TTC CCG ACC GAG CCG AAC AGC TCC AGC CGC GCC ATC CAC Leu Glu Gly Phe Pro Thr Glu Pro Asn Ser Ser Ser Arg Ala Ile His 85 90 95	288
GTG CAC CAG TTC GGG GAC CTG AGC CAG GGC TGC GAG TCC ACC GGG CCC Val His Gln Phe Gly Asp Leu Ser Gln Gly Cys Glu Ser Thr Gly Pro 100 105 110	336
CAC TAC AAC CCG CTG GCC GTG CCG CAC CCG CAG CAC CCG GGC GAC TTC His Tyr Asn Pro Leu Ala Val Pro His Pro Gln His Pro Gly Asp Phe 115 120 125	384
GGC AAC TTC GCG GTC CGC GAC GGC AGC CTC TGG AGG TAC CGC GCC GGC Gly Asn Phe Ala Val Arg Asp Gly Ser Leu Trp Arg Tyr Arg Ala Gly 130 135 140	432
CTG GCC GCC TCG CTC GCG GGC CCG CAC TCC ATC GTG GGC CGG GCC GTG Leu Ala Ala Ser Leu Ala Gly Pro His Ser Ile Val Gly Arg Ala Val 145 150 155 160	480
GTC GTC CAC GCT GGC GAG GAC GAC CTG GGC CGC GGC GGC AAC CAG GCC Val Val His Ala Gly Glu Asp Asp Leu Gly Arg Gly Gly Asn Gln Ala 165 170 175	528
AGC GTG GAG AAC GGG AAC GCG GGC CGG CGG CTG GCC TGC TGC GTG GTG Ser Val Glu Asn Gly Asn Ala Gly Arg Arg Leu Ala Cys Cys Val Val 180 185 190	576
GGC GTG TGC GGG CCC GGG CTC TGG GAG CGC CAG GCG CGG GAG CAC TCA Gly Val Cys Gly Pro Gly Leu Trp Glu Arg Gln Ala Arg Glu His Ser 195 200 205	624
GAG CGC AAG AAG CGG CGG CGC GAG AGC GAG TGC AAG GCC GCC TGA Glu Arg Lys Lys Arg Arg Arg Glu Ser Glu Cys Lys Ala Ala 210 215 220	669

## (2) INFORMATION FOR SEQ ID NO: 2:

## (i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 222 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

71

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 2:

```

Trp Thr Gly Glu Asp Ser Ala Glu Pro Asn Ser Asp Ser Ala Glu Trp
 1           5           10           15

Ile Arg Asp Met Tyr Ala Lys Val Thr Glu Ile Trp Gln Glu Val Met
      20           25           30

Gln Arg Arg Asp Asp Asp Gly Thr Leu His Ala Ala Cys Gln Val Gln
      35           40           45

Pro Ser Ala Thr Leu Asp Ala Ala Gln Pro Arg Val Thr Gly Val Val
      50           55           60

Leu Phe Arg Gln Leu Ala Pro Arg Ala Lys Leu Asp Ala Phe Phe Ala
      65           70           75           80

Leu Glu Gly Phe Pro Thr Glu Pro Asn Ser Ser Ser Arg Ala Ile His
      85           90           95

Val His Gln Phe Gly Asp Leu Ser Gln Gly Cys Glu Ser Thr Gly Pro
      100          105          110

His Tyr Asn Pro Leu Ala Val Pro His Pro Gln His Pro Gly Asp Phe
      115          120          125

Gly Asn Phe Ala Val Arg Asp Gly Ser Leu Trp Arg Tyr Arg Ala Gly
      130          135          140

Leu Ala Ala Ser Leu Ala Gly Pro His Ser Ile Val Gly Arg Ala Val
      145          150          155          160

Val Val His Ala Gly Glu Asp Asp Leu Gly Arg Gly Gly Asn Gln Ala
      165          170          175

Ser Val Glu Asn Gly Asn Ala Gly Arg Arg Leu Ala Cys Cys Val Val
      180          185          190

Gly Val Cys Gly Pro Gly Leu Trp Glu Arg Gln Ala Arg Glu His Ser
      195          200          205

Glu Arg Lys Lys Arg Arg Arg Glu Ser Glu Cys Lys Ala Ala
      210          215          220

```

(2) INFORMATION FOR SEQ ID NO: 3:

- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 25 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 3:

CTGTGTGGCC AAGAAGGAAG TTTGT

25

(2) INFORMATION FOR SEQ ID NO: 4:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 20 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 4:

GCCTGAGGAT GAGCCAAGTG

20

(2) INFORMATION FOR SEQ ID NO: 5:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 27 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 5:

GTCCAGCGTG GCCGACGGCT GCACCTG

27

(2) INFORMATION FOR SEQ ID NO: 6:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 51 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(v) FRAGMENT TYPE: C-terminal

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 1..51



73

(D) OTHER INFORMATION: /product= "C-terminal of native EC-SOD"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 6:

CAC	TCA	GAG	CGC	AAG	AAG	CGG	CGG	CGC	GAG	AGC	GAG	TGC	AAG	GCC	GCC	48
His	Ser	Glu	Arg	Lys	Lys	Arg	Arg	Arg	Glu	Ser	Glu	Cys	Lys	Ala	Ala	
1				5					10					15		
TGA																51

(2) INFORMATION FOR SEQ ID NO: 7:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 16 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 7:

His	Ser	Glu	Arg	Lys	Lys	Arg	Arg	Arg	Glu	Ser	Glu	Cys	Lys	Ala	Ala
1				5					10					15	

(2) INFORMATION FOR SEQ ID NO: 8:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 51 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(v) FRAGMENT TYPE: C-terminal

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 1..12
- (D) OTHER INFORMATION: /product= "C-terminal of variant T209"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 8:

CAC	TCA	GAG	TGAAAGAAGC	GGCGGCGCGA	GAGCGAGTGC	AAGGCCGCCT	GA	51
His	Ser	Glu						
1								

## (2) INFORMATION FOR SEQ ID NO: 9:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 3 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 9:

His Ser Glu

1

## (2) INFORMATION FOR SEQ ID NO: 10:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 51 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(v) FRAGMENT TYPE: C-terminal

## (ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 1..24
- (D) OTHER INFORMATION: /product= "C-terminal of variant T213"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 10:

CAC TCA GAG CGC AAG AAG CGG TGACGCGAGA GCGAGTGCAA GGCCGCCTGA  
His Ser Glu Arg Lys Lys Arg

51

1

5

## (2) INFORMATION FOR SEQ ID NO: 11:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 7 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 11:

His Ser Glu Arg Lys Lys Arg

1

5

(2) INFORMATION FOR SEQ ID NO: 12:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 51 base pairs  
(B) TYPE: nucleic acid  
(C) STRANDEDNESS: single  
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(v) FRAGMENT TYPE: C-terminal

(ix) FEATURE:

- (A) NAME/KEY: CDS  
(B) LOCATION: 1..30  
(D) OTHER INFORMATION: /product= "C-terminal of variant T215"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 12:

CAC TCA GAG CGC AAG AAG CGG CGG CGC TAGAGCGAGT GCAAGGCCGC	47
His Ser Glu Arg Lys Lys Arg Arg Arg	
1 5 10	
CTGA	51

## (2) INFORMATION FOR SEQ ID NO: 13:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 9 amino acids  
(B) TYPE: amino acid  
(D) TOPOLOGY: linear

## (ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 13:

His Ser Glu Arg Lys Lys Arg Arg Arg  
1 5

(2) INFORMATION FOR SEQ ID NO: 14:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 51 base pairs  
(B) TYPE: nucleic acid  
(C) STRANDEDNESS: single  
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(v) FRAGMENT TYPE: C-terminal

76

## (ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 1..33
- (D) OTHER INFORMATION: /product= "C-terminal of variant T216"

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 14:

CAC TCA GAG CGC AAG AAG CGG CGG CGC GAG TGAGAGTGCA AGGCCGCCTG A 51  
 His Ser Glu Arg Lys Lys Arg Arg Arg Glu  
 1 5 10

## (2) INFORMATION FOR SEQ ID NO: 15:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 10 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

## (ii) MOLECULE TYPE: protein

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 15:

His Ser Glu Arg Lys Lys Arg Arg Arg Glu  
 1 5 10

## (2) INFORMATION FOR SEQ ID NO: 16:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 51 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

## (ii) MOLECULE TYPE: cDNA

## (v) FRAGMENT TYPE: C-terminal

## (ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 1..51
- (D) OTHER INFORMATION: /product= "C-terminal of variant SA216"

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 16:

CAC TCA GAG CGC AAG AAG CGG CGG CGC GCG AGC GAG TGC AAG GCC GCC 48  
 His Ser Glu Arg Lys Lys Arg Arg Arg Ala Ser Glu Cys Lys Ala Ala  
 1 5 10 15

TGA

51

## (2) INFORMATION FOR SEQ ID NO: 17:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 16 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

## (ii) MOLECULE TYPE: protein

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 17:

His Ser Glu Arg Lys Lys Arg Arg Arg Ala Ser Glu Cys Lys Ala Ala  
1 5 10 15

## (2) INFORMATION FOR SEQ ID NO: 18:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 51 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

## (ii) MOLECULE TYPE: cDNA

## (v) FRAGMENT TYPE: C-terminal

## (ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 1..51
- (D) OTHER INFORMATION: /product= "C-terminal of variant SA219"

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 18:

CAC TCA GAG CGC AAG AAG CGG CGG CGC GAG AGC GAG GCC AAG GCC GCC 48  
His Ser Glu Arg Lys Lys Arg Arg Arg Glu Ser Glu Ala Lys Ala Ala  
1 5 10 15  
TGA 51

## (2) INFORMATION FOR SEQ ID NO: 19:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 16 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

## (ii) MOLECULE TYPE: protein

78

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 19:

His Ser Glu Arg Lys Lys Arg Arg Arg Glu Ser Glu Ala Lys Ala Ala  
 1 5 10 15

(2) INFORMATION FOR SEQ ID NO: 20:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 51 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(v) FRAGMENT TYPE: C-terminal

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 1..51
- (D) OTHER INFORMATION: /product= "C-terminal of variant SA220"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 20:

CAC TCA GAG CGC AAG AAG CGG CGG CGC GAG AGC GAG TGC GCG GCC GCC 48  
 His Ser Glu Arg Lys Lys Arg Arg Arg Glu Ser Glu Cys Ala Ala Ala  
 1 5 10 15  
 TGA 51

(2) INFORMATION FOR SEQ ID NO: 21:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 16 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 21:

His Ser Glu Arg Lys Lys Arg Arg Arg Glu Ser Glu Cys Ala Ala Ala  
 1 5 10 15

(2) INFORMATION FOR SEQ ID NO: 22:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 51 base pairs
- (B) TYPE: nucleic acid

(C) STRANDEDNESS: single  
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(v) FRAGMENT TYPE: C-terminal

(ix) **FEATURE:**

(A) NAME/KEY: CDS

(B) LOCATION: 1..51

(D) OTHER INFORMATION: /product= "C-terminal of variant  
SA211-213"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 22:

CAC	TCA	GAG	CGC	GCG	GCG	GCG	CGG	CGC	GAG	AGC	GAG	TGC	AAG	GCC	GCC		48
His	Ser	Glu	Arg	Ala	Ala	Ala	Arg	Arg	Glu	Ser	Glu	Cys	Lys	Ala	Ala		
1				5					10					15			
 TGA																	51

(2) INFORMATION FOR SEQ ID NO: 23:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 16 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 23:

His Ser Glu Arg Ala Ala Ala Arg Arg Glu Ser Glu Cys Lys Ala Ala  
1 5 10 15

(2) INFORMATION FOR SEQ ID NO: 24:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 51 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(v) FRAGMENT TYPE: C-terminal

(ix) **FEATURE:**

(A) NAME/KEY: CDS

(B) LOCATION: 1..51

(D) OTHER INFORMATION: /product= "C-terminal of variant  
SA216/218"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 24:

[illegible]

(2) INFORMATION FOR SEQ ID NO: 25:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 16 amino acids  
(B) TYPE: amino acid  
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 25:

His Ser Glu Arg Lys Lys Arg Arg Arg Ala Ser Ala Cys Lys Ala Ala  
1 5 10 15

(2) INFORMATION FOR SEQ ID NO: 26:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 51 base pairs  
(B) TYPE: nucleic acid  
(C) STRANDEDNESS: single  
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(v) FRAGMENT TYPE: C-terminal

(ix) **FEATURE:**

- (A) NAME/KEY: CDS  
(B) LOCATION: 1..51  
(D) OTHER INFORMATION: /product= "C-terminal of variant  
SA216/220"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 26:

CAC	TCA	GAG	CGC	AAG	AAG	CGG	CGG	CGC	GCG	AGC	GAG	TGC	GCG	GCC	GCC	48
His	Ser	Glu	Arg	Lys	Lys	Arg	Arg	Arg	Ala	Ser	Glu	Cys	Ala	Ala	Ala	
1				5					10					15		
TGA																51



## (2) INFORMATION FOR SEQ ID NO: 27:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 16 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

## (ii) MOLECULE TYPE: protein

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 27:

His Ser Glu Arg Lys Lys Arg Arg Arg Ala Ser Glu Cys Ala Ala Ala  
1 5 10 15

## (2) INFORMATION FOR SEQ ID NO: 28:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 51 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

## (ii) MOLECULE TYPE: cDNA

## (v) FRAGMENT TYPE: C-terminal

## (ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 1..33
- (D) OTHER INFORMATION: /product= "C-terminal of variant SAT216"

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 28:

CAC TCA GAG CGC AAG AAG CGG CGG CGC GCG TGAGAGTGCA AGGCCGCCTG A 51  
His Ser Glu Arg Lys Lys Arg Arg Arg Ala  
1 5 10

## (2) INFORMATION FOR SEQ ID NO: 29:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 10 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

## (ii) MOLECULE TYPE: protein

82

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 29:

His Ser Glu Arg Lys Lys Arg Arg Arg Ala  
1 5 10

(2) INFORMATION FOR SEQ ID NO: 30:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 51 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(v) FRAGMENT TYPE: C-terminal

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 1..33
- (D) OTHER INFORMATION: /product= "C-terminal of variant SRT216"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 30:

CAC TCA GAG CGC AAG AAG CGG CGG CGC CGG TGAGAGTGCA AGGCCGCCTG A 51  
His Ser Glu Arg Lys Lys Arg Arg Arg Arg  
1 5 10

(2) INFORMATION FOR SEQ ID NO: 31:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 10 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein


(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 31:

His Ser Glu Arg Lys Lys Arg Arg Arg Arg  
1 5 10

## INDICATIONS RELATING TO A DEPOSITED MICROORGANISM

(PCT Rule 13bis)

<b>A.</b> The indications made below relate to the microorganism referred to in the description on page <u>21</u> , line <u>24</u>	
<b>B. IDENTIFICATION OF DEPOSIT</b> <span style="float: right;">Further deposits are identified on an additional sheet <input type="checkbox"/></span>	
Name of depositary institution  Deutsche Sammlung von Mikroorganismen und Zellkulturen GmbH (DSMZ)	
Address of depositary institution (including postal code and country) Maschroder Weg 1b D-38124 Braunschweig Federal Republic of Germany	
Date of deposit 7 June 1993	Accession Number DSMZ 8335
<b>C. ADDITIONAL INDICATIONS</b> (leave blank if not applicable) <span style="float: right;">This information is continued on an additional sheet <input type="checkbox"/></span>	
As regards the respective Patent Offices of the respective designated states, the applicant requests that a sample of the deposited microorganisms only be made available to an expert nominated by the requester until the date on which the patent is granted or the date on which the application has been refused or withdrawn or is deemed to be withdrawn	
<b>D. DESIGNATED STATES FOR WHICH INDICATIONS ARE MADE</b> (if the indications are not for all designated States)	
<b>E. SEPARATE FURNISHING OF INDICATIONS</b> (leave blank if not applicable)	
The indications listed below will be submitted to the International Bureau later (specify the general nature of the indications e.g., "Accession Number of Deposit")	

For receiving Office use only	
<input checked="" type="checkbox"/> This sheet was received with the international application	
Authorized officer  A. LORIS	

For International Bureau use only	
<input type="checkbox"/> This sheet was received by the International Bureau on:	
Authorized officer	

## CLAIMS

1. A mammalian expression system comprising a DNA sequence encoding a polypeptide having the amino acid sequence shown in SEQ ID NO:2 or variant thereof which has a biological  
5 activity of human EC-SOD, said DNA sequence is combined with regulatory element of a gene encoding a milk protein of a mammal so as to form a hybrid gene which is expressible in the mammary gland of an adult female of a non-human mammal harbouring said hybrid gene so that the polypeptide encoded  
10 by the DNA sequence is produced when the hybrid gene is expressed.
2. An expression system according to claim 1, wherein the gene encoding a milk protein is selected from the group consisting of whey acidic protein (WAP) genes,  $\beta$ -  
15 lactoglobulin genes and casein genes.
3. An expression system according to claim 1, wherein the variant of the polypeptide is at least 85% homologous with the amino acid sequence shown in SEQ ID NO:2.
4. An expression system according to claim 1, wherein the DNA  
20 sequence encoding the polypeptide is one which hybridizes with the DNA sequence shown in SEQ ID NO:1 or with a part thereof.
5. An expression system according to claim 4, wherein the DNA sequence encoding the polypeptide contains at least one  
25 intron sequence.
6. An expression system according to claim 1, wherein the DNA sequence encoding the polypeptide is preceded by a DNA sequence encoding a signal peptide.
7. An expression system according to claim 6, wherein the  
30 signal peptide is selected from the group consisting of signal peptides associated with the DNA sequence encoding

human EC-SOD and signal peptides associated with the milk protein.

8. An expression system according to claim 1, wherein the non-human mammal is selected from the group consisting of rabbits, mice, rats, goats, sheep, pigs, lama, camels and bovine species.

9. An expression system according to claim 1, wherein the variant has a reduced heparin affinity as compared to human EC-SOD type C.

10. An expression system according to claim 9, wherein the variant is selected from the group consisting of variants T216, T215, T213, T209, SAT216, SRT216, SA219, SA220 and SA211-213.

11. An expression system according to claim 1, wherein the variant has an increased heparin affinity as compared to human EC-SOD type C.

12. An expression system according to claim 11, wherein the variant is selected from the group consisting of variants SA216, SA216/218 and SA216/220.

13. A DNA fragment comprising a DNA sequence encoding a polypeptide having the amino acid sequence shown in SEQ ID NO:2 or variant thereof which has a biological activity of human EC-SOD, said DNA sequence comprising at least one intron sequence.

14. A DNA fragment according to claim 13, wherein the DNA sequence differs from the DNA sequence shown in SEQ ID NO:1 in that at least one nucleotide has been deleted, substituted or modified or at least one additional nucleotide has been inserted.

15. A DNA fragment according to claim 14, wherein the DNA sequence

5       1) hybridizes with the DNA sequence shown in SEQ ID NO:1 or a specific part thereof, preferably under stringent hybridization conditions or

      2) encodes a polypeptide, the amino acid sequence of which is at least 85% homologous with the amino acid sequence shown in SEQ ID NO:2, or

10       3) constitutes an effective subsequence of said DNA sequence.

16. A DNA fragment according to claim 14, wherein the DNA sequence encodes a variant selected from the group consisting of variants T216, T215, T213, T209, SAT216, SRT216, SA219, SA220, SA211-213, SA216, SA216/218 and SA216/220.

15 17. A DNA fragment according to claim 14, wherein the DNA sequence originates from cDNA, genomic DNA, mixed cDNA and genomic DNA, mixed cDNA and synthetic DNA, or mixed genomic DNA and synthetic DNA.

20 18. A hybrid gene comprising a DNA sequence encoding a polypeptide having the amino acid sequence shown in SEQ ID NO:2 or a variant thereof which has a biological activity of human EC-SOD, said DNA sequence being combined with a regulatory element of a gene encoding a milk protein of a mammal.

25 19. A hybrid gene according to claim 18, wherein the gene encoding the milk protein is selected from the group consisting of whey acidic protein (WAP) genes,  $\beta$ -lactoglobulin genes or casein genes.

20. A hybrid gene according to claim 18 or claim 19, wherein the variant is selected from the group consisting of variants T216, T215, T213, T209, SAT216, SRT216, SA219, SA220, SA211-213, SA216, SA216/218 and SA216/220.

5 21. A hybrid gene according to claim 18, wherein the DNA sequence comprises at least one intron sequence.

22. A replicable expression vector which carries a DNA sequence encoding a polypeptide having the amino acid sequence shown in SEQ ID NO:2 or variant thereof which has a  
10 biological activity of human EC-SOD comprising the DNA sequence shown in SEQ ID NO:1 or a variant thereof, said DNA sequence comprising at least one intron sequence and is capable of mediating the expression of said DNA sequence.

23. A replicable expression vector according to claim 22,  
15 wherein the DNA sequence differs from the DNA sequence shown in SEQ ID NO:1 in that at least one nucleotide has been deleted, substituted or modified or at least one additional nucleotide has been inserted.

24. A replicable expression vector according to claim 22,  
20 wherein the DNA sequence is as defined in any of claims 13-21.

25. A replicable expression vector according to claim 22 designated pS 172 which has been deposited 7 June 1993 with the collection of Deutsche Sammlung von Mikroorganismen und  
25 Zellkulturen GmbH (DSMZ) under the accession number DSM 8335 in accordance with the provisions of the Budapest Treaty, and expression vectors expressing DNA sequences which differ from the DNA sequences of the said deposited expression vectors.

26. A replicable expression vector according to claim 25,  
30 wherein the DNA sequence expressed is one which differs from the DNA sequence of the deposited vector in that at least one nucleotide has been deleted, substituted or modified or at

least one additional nucleotide has been inserted so as to result in a DNA sequence which encodes a polypeptide having a biological activity of EC-SOD.

27. A cell harbouring a vector as defined in any of claims  
5 22-26.

28. A cell according to claim 27 which is selected from a group consisting of a unicellular eukaryotic cell or a cell derived from a multicellular organism.

29. A cell according to claim 28, which is derived from a  
10 non-human mammal.

30. A method for producing a polypeptide having the amino acid sequence shown in SEQ ID NO:2 or variant thereof which has a biological activity of human EC-SOD, said method comprising introducing at least one expression system as claimed  
15 in any of claims 1-12 into the genome of a non-human mammal in such a way that the DNA encoding the polypeptide is expressed in a mammary gland of the non-human mammal, collecting the milk secreted from the gland, and recovering the recombinant polypeptide from the milk, optionally followed by  
20 purification.

31. A method for producing a transgenic non-human mammal capable of expressing a polypeptide having the amino acid sequence SEQ ID NO:2 or variant thereof which has a biological activity of human EC-SOD said method comprising chromosomally incorporating at least one DNA sequence encoding the  
25 polypeptide into the genome of a non-human mammal.

32. A method according to claim 31, comprising injecting at least one expression system as defined in any of claims 1-12 or subsequence thereof into a fertilized egg or a cell of an  
30 embryo of a mammal so as to incorporate the expression system into the germline of the mammal and developing the resulting



injected fertilized egg or embryo into an adult female mammal.

33. A method according to any of claims 30-32, wherein 1-10  
5 copies of the expression system or a subsequence thereof are injected.

34. A method according to claim 33, wherein 1-7 copies of the expression system or a subsequence thereof are injected.

35. A method according to claim 33 or 34, wherein the expression systems injected comprises at least two different DNA  
10 sequences encoding two different variants of EC-SOD.

36. A transgenic non-human mammal whose germ cells and somatic cells contain a DNA sequence encoding a polypeptide having the amino acid sequence SEQ ID NO:2 or variant thereof said polypeptide or variant having a biological activity of  
15 human EC-SOD as a result of chromosomal incorporation into the non-human mammalian genome, or into the genome of an ancestor of said non-human mammal.

37. A transgenic non-human mammal according to claim 36, wherein the DNA sequence is a DNA sequence as defined in any  
20 of claims 13-21..

38. A transgenic non-human mammal as claimed in claim 37, wherein the DNA sequence is present in a milk protein gene of the mammal.

39. A transgenic non-human mammal according to claim 36,  
25 which is prepared by the method defined in any of claims 31-34.

40. A transgenic non-human mammal according to any of claims 36-39, which is selected from the group consisting of rabbits, mice, rats, goats, sheep, pigs, lama, camels and  
30 bovine species.

41. Milk from a transgenic non-human mammal comprising a polypeptide having the amino acid sequence SEQ ID NO:2 or a variant thereof which has a biological activity of human EC-SOD together with milk constituents.

- 5 42. Milk according to claim 41, wherein the milk is obtained from a transgenic mammal as defined in any of claims 36-40.

1/8

	210	215	220
Native	HisSerGluArgLysLysArgArgArgGluSerGluCysLysAlaAla***		
	CACTCAGAGCGCAAGAAGCGGCGGCGCGAGAGCGAGTGCAAGGCCGCCTGA		
T209	HisSerGlu***		
	CACTCAGAGTGAAAGAAGCGGCGGCGCGAGAGCGAGTGCAAGGCCGCCTGA		
T213	HisSerGluArgLysLysArg***		
	CACTCAGAGCGCAAGAAGCGGTGACGCGAGAGCGAGTGCAAGGCCGCCTGA		
T215	HisSerGluArgLysLysArgArgArg***		
	CACTCAGAGCGCAAGAAGCGGCGGCGCTAGAGCGAGTGCAAGGCCGCCTGA		
T216	HisSerGluArgLysLysArgArgArgGlu***		
	CACTCAGAGCGCAAGAAGCGGCGGCGCGAGTGAGAGTGCAAGGCCGCCTGA		
SA216	HisSerGluArgLysLysArgArgArgAlaSerGluCysLysAlaAla***		
	CACTCAGAGCGCAAGAAGCGGCGGCGCGAGCGAGTGCAAGGCCGCCTGA		
SA219	HisSerGluArgLysLysArgArgArgGluSerGluAlaLysAlaAla***		
	CACTCAGAGCGCAAGAAGCGGCGGCGCGAGAGCGAGGCCAAGGCCGCCTGA		
SA220	HisSerGluArgLysLysArgArgArgGluSerGluCysAlaAlaAla***		
	CACTCAGAGCGCAAGAAGCGGCGGCGCGAGAGCGAGTGCGCGGCCGCCTGA		
SA211-213	HisSerGluArgAlaAlaAlaArgArgGluSerGluCysLysAlaAla***		
	CACTCAGAGCGCGCGGCGGCGGCGCGAGAGCGAGTGCAAGGCCGCCTGA		
SA216/218	HisSerGluArgLysLysArgArgArgAlaSerAlaCysLysAlaAla***		
	CACTCAGAGCGCAAGAAGCGGCGGCGCGAGAGCGCGTGCAAGGCCGCCTGA		
SA216/220	HisSerGluArgLysLysArgArgArgAlaSerGluCysAlaAlaAla***		
	CACTCAGAGCGCAAGAAGCGGCGGCGCGAGAGCGAGTGCGCGGCCGCCTGA		
SAT216	HisSerGluArgLysLysArgArgArgAla***		
	CACTCAGAGCGCAAGAAGCGGCGGCGCGCGTGAGAGTGCAAGGCCGCCTGA		
SRT216	HisSerGluArgLysLysArgArgArgArg***		
	CACTCAGAGCGCAAGAAGCGGCGGCGCCGGTGAGAGTGCAAGGCCGCCTGA		

Fig. 1

2/8

Fig. 2A

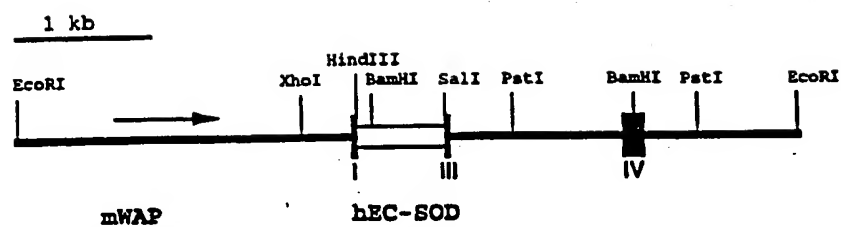
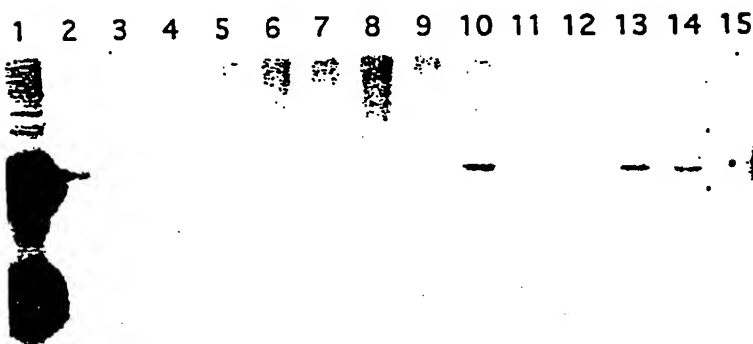


Fig. 2B



3/8

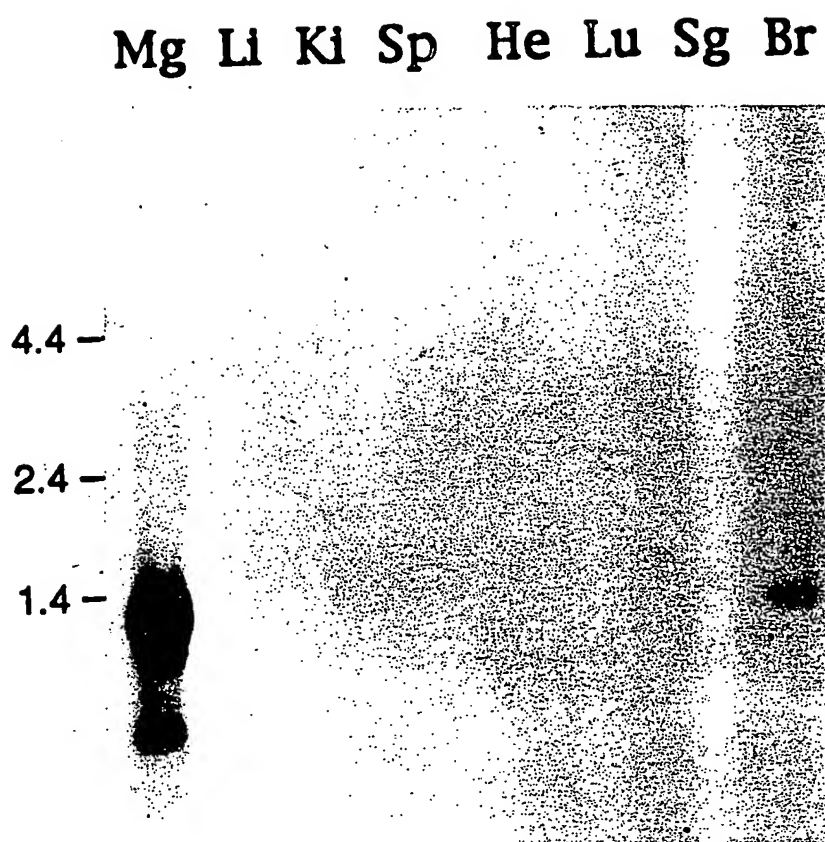


Fig. 3A

4/8

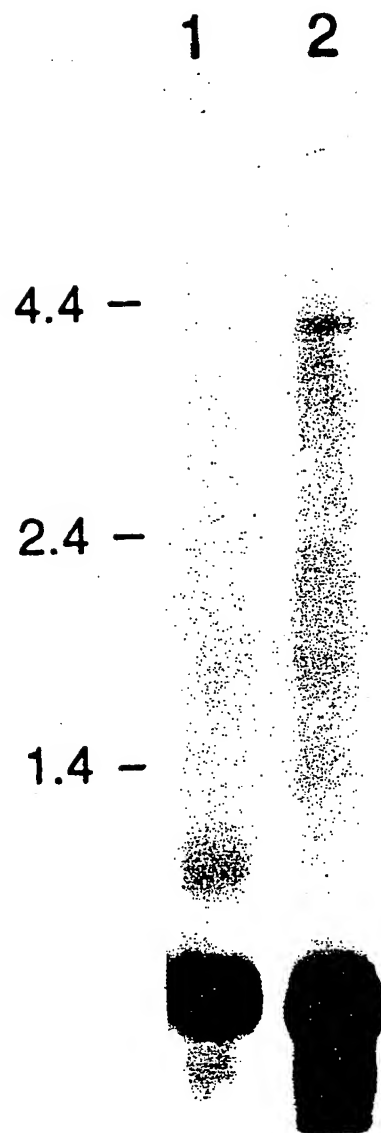


Fig. 3B

5/8

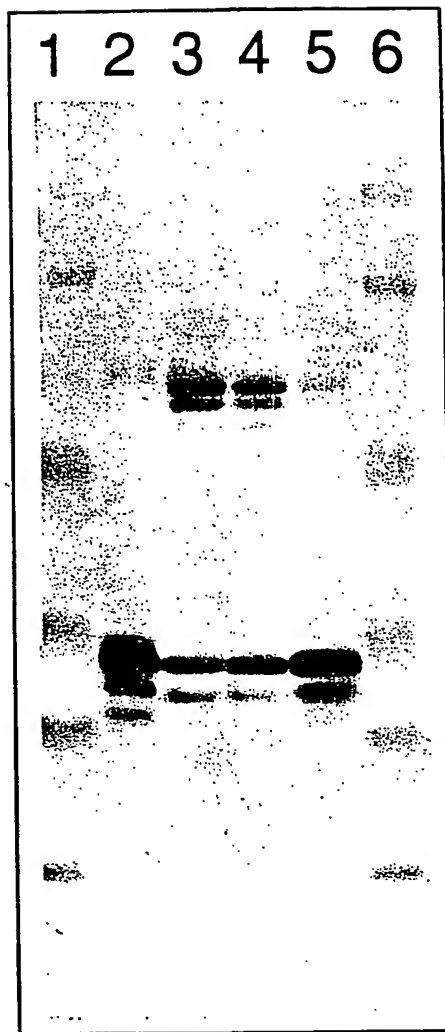


Fig. 4

6/8

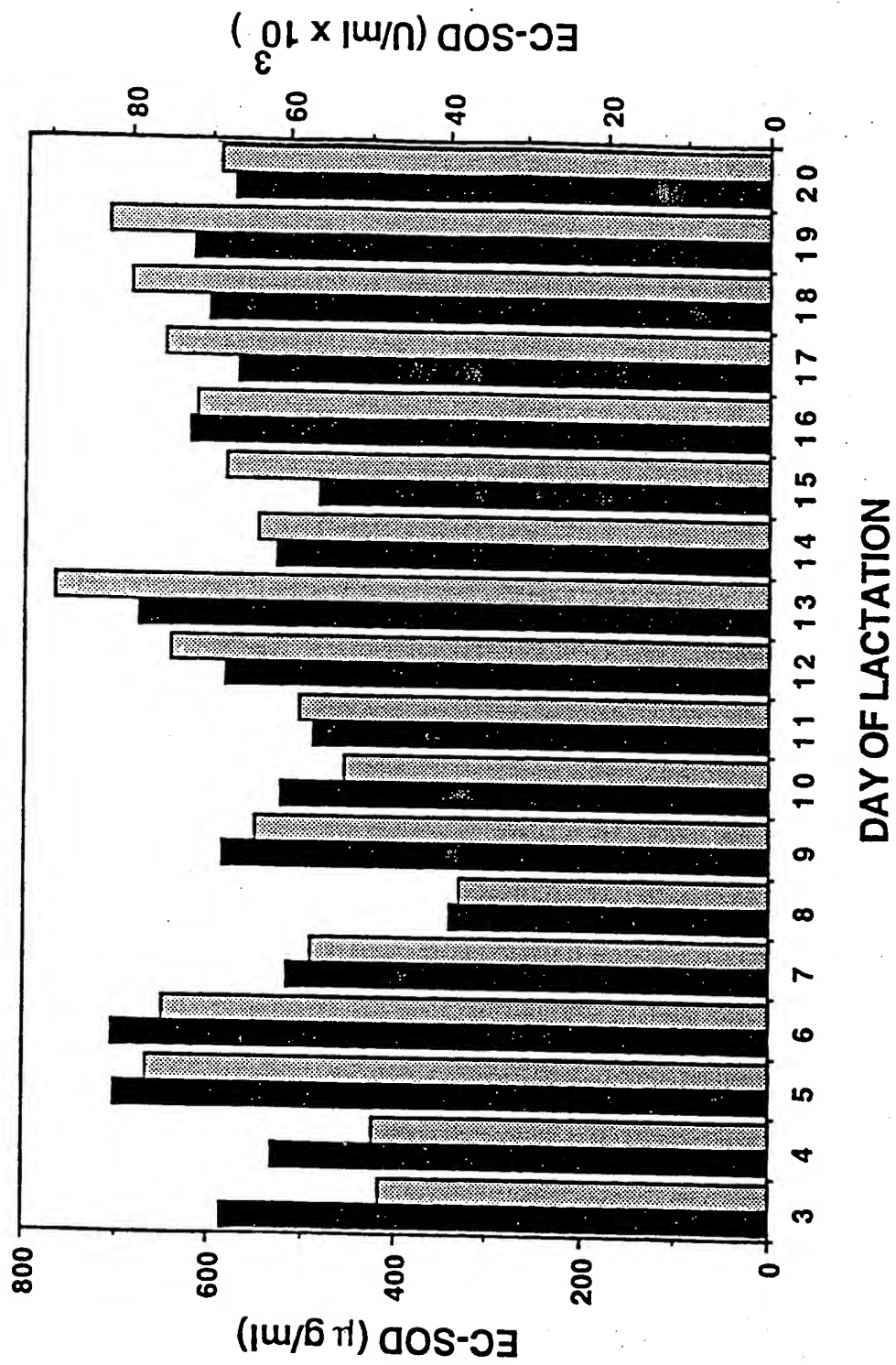


Fig. 5



7/8

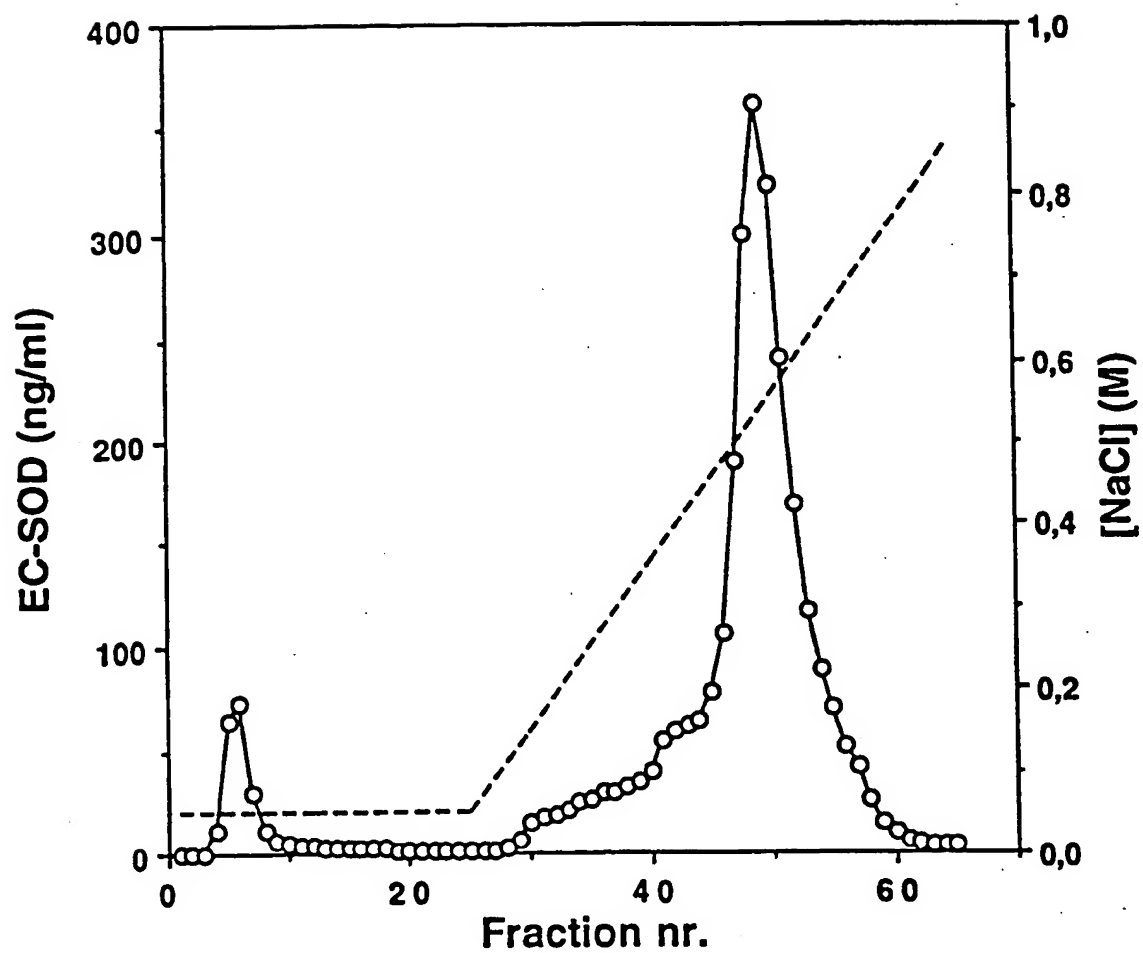


Fig. 6

8/8

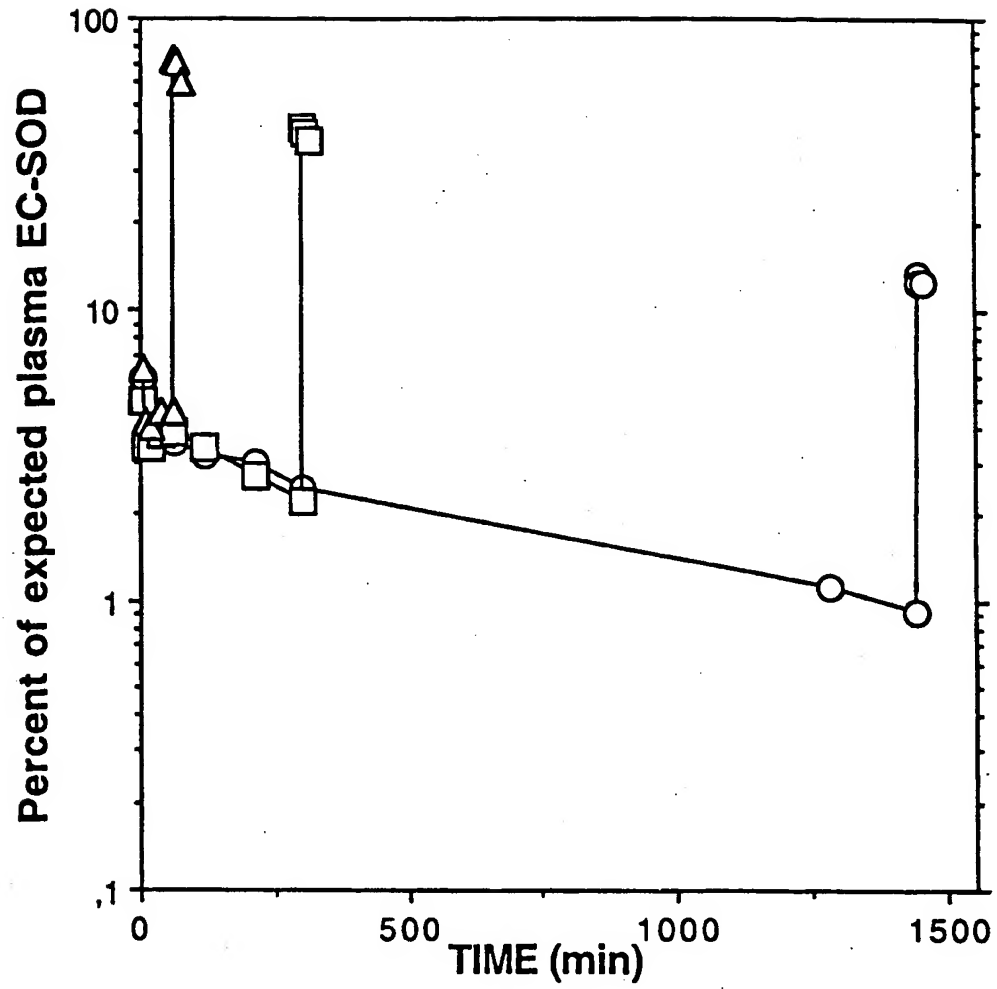


Fig. 7

## A. CLASSIFICATION OF SUBJECT MATTER

IPC 5 C12N15/00 A01K67/027 C12N9/02 C12N15/53 C12N5/10  
A23C9/20

According to International Patent Classification (IPC) or to both national classification and IPC

## B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 5 A01K C12N A23C

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

## C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	PROCEEDINGS OF THE NATIONAL ACADEMY OF SCIENCES OF USA. vol. 89 , October 1992 , WASHINGTON US pages 9715 - 9719 OURY, T.D. ET AL. 'Extracellular superoxide dismutase' cited in the application see the whole document ---	1-42
P,Y	JOURNAL OF BIOLOGICAL CHEMISTRY. (MICROFILMS) vol. 268, no. 21 , 25 July 1993 , BALTIMORE, MD US pages 15394 - 15398 OURY, T.D. ET AL. 'Cold-induced brain edema in mice' see the whole document --- -/--	1-42

☒ Further documents are listed in the continuation of box C.

☒ Patent family members are listed in annex.

## \* Special categories of cited documents :

- \*A\* document defining the general state of the art which is not considered to be of particular relevance
- \*E\* earlier document but published on or after the international filing date
- \*L\* document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)
- \*O\* document referring to an oral disclosure, use, exhibition or other means
- \*P\* document published prior to the international filing date but later than the priority date claimed

\*T\* later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

\*X\* document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

\*Y\* document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.

\*Z\* document member of the same patent family

Date of the actual completion of the international search

31 August 1994

Date of mailing of the international search report

- 9. 09. 94

Name and mailing address of the ISA

European Patent Office, P.B. 5818 Patentlaan 2  
NL - 2280 HV Rijswijk  
Tel. (+ 31-70) 340-2040, Tx. 31 651 epo nl,  
Fax (+ 31-70) 340-3016

Authorized officer

Chambonnet, F

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT		
Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	<p>PROTEIN EXPRESSION AND PURIFICATION vol. 1 , 1990 pages 3 - 8 HENNIGHAUSEN, L. 'The mammary gland as a bioreactor: production of foreign proteins in milk' cited in the application see the whole document ---</p>	1-42
X	<p>WO,A,88 10118 (BIOGEN, N.V.) 29 December 1988 cited in the application see the whole document ---</p>	13
Y	<p>WO,A,88 10118 (BIOGEN, N.V.) 29 December 1988 cited in the application see the whole document ---</p>	1-12, 18-42
Y	<p>WO,A,90 05188 (PHARMACEUTICAL PROTEINS LIMITED) 17 May 1990 see the whole document ---</p>	1-42
X	<p>WO,A,87 01387 (SYN-TEK AB) 12 March 1987 cited in the application see the whole document ---</p>	13-17
Y	<p>WO,A,87 01387 (SYN-TEK AB) 12 March 1987 cited in the application see the whole document ---</p>	1-42
Y	<p>WO,A,91 04315 (SYMBICOM) 4 April 1991 see the whole document -----</p>	1-42

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
WO-A-8810118	29-12-88	US-A- 4873316	10-10-89
		EP-A- 0347431	27-12-89
		JP-T- 2500798	22-03-90
-----			
WO-A-9005188	17-05-90	AU-B- 628101	10-09-92
		AU-A- 4494389	28-05-90
		EP-A- 0396699	14-11-90
		JP-T- 3505674	12-12-91
-----			
WO-A-8701387	12-03-87	AU-B- 598756	05-07-90
		AU-A- 6370886	24-03-87
		DE-A- 3682505	19-12-91
		EP-A, B 0236385	16-09-87
		JP-T- 63501473	09-06-88
		SU-A- 1779263	30-11-92
		US-A- 5130245	14-07-92
		US-A- 5248603	28-09-93
-----			
WO-A-9104315	04-04-91	AU-A- 6444190	18-04-91
		EP-A- 0593435	27-04-94
-----			

**THIS PAGE BLANK (USPTO)**